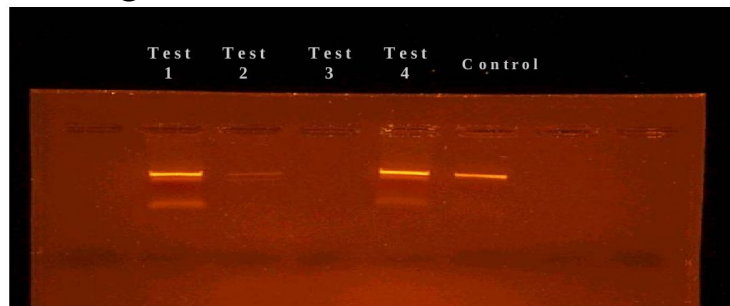
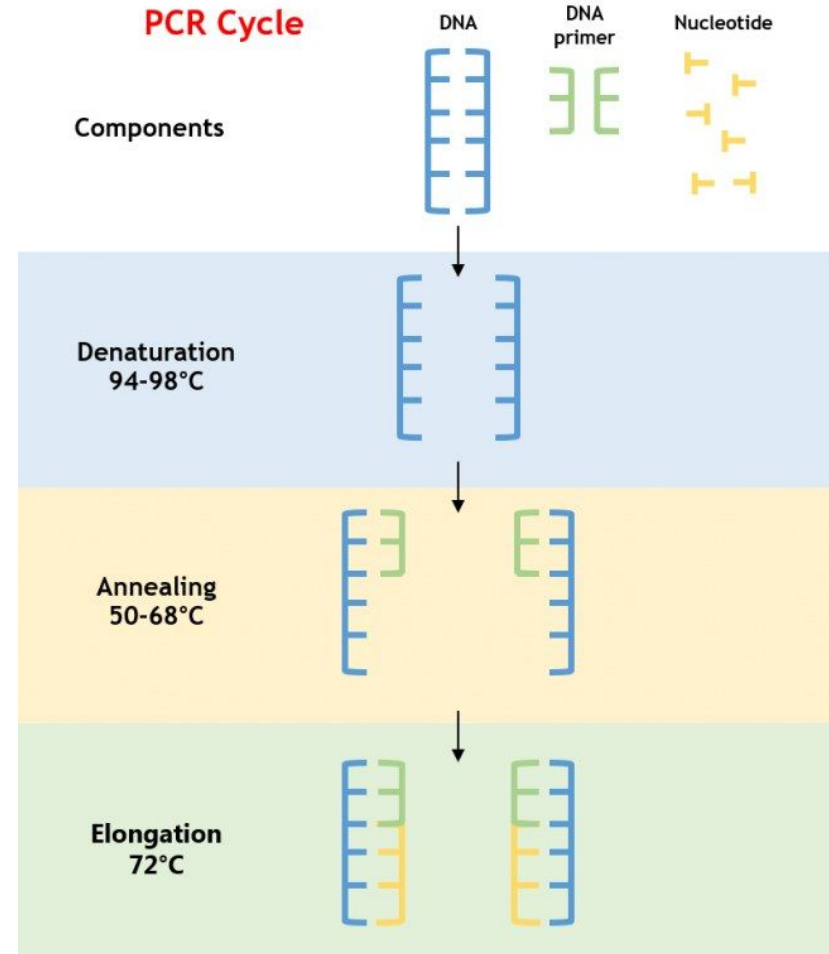
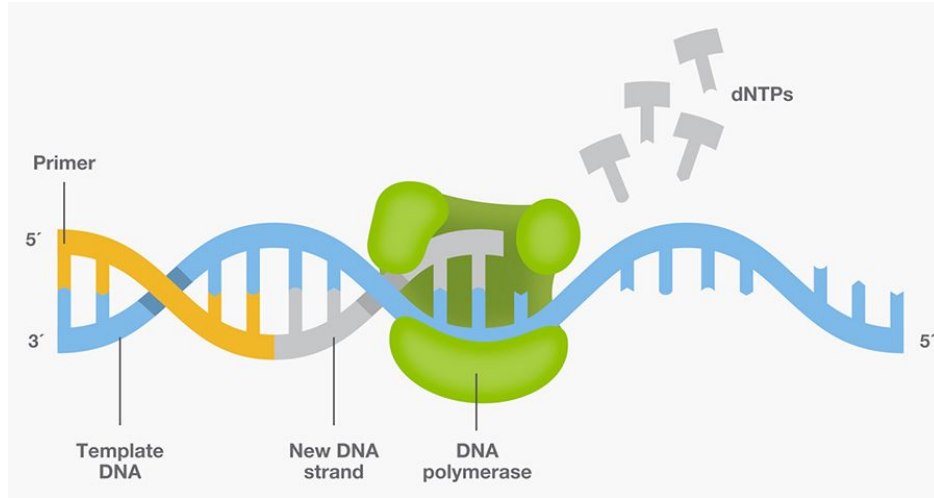


My Kitchen Table PCR Sophomore Year of High School



PCR Primer



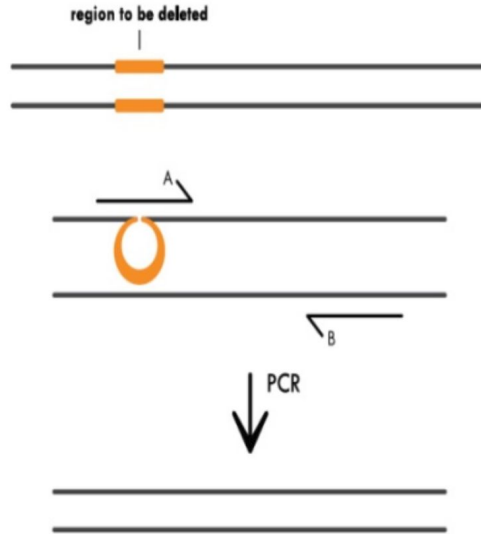
Primer-Defined Changes to the PCR Copies

-

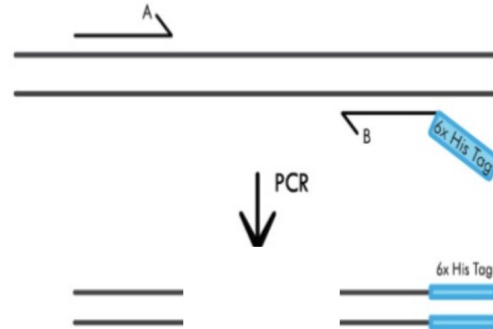
+

Δ

PCR for Deletions



PCR for Terminal Additions



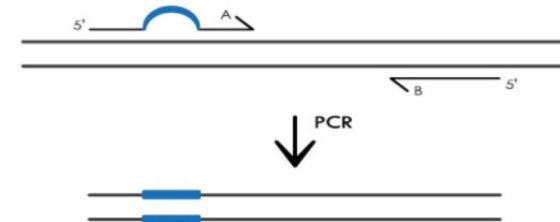
PCR for Base Substitutions.

Existing sequence

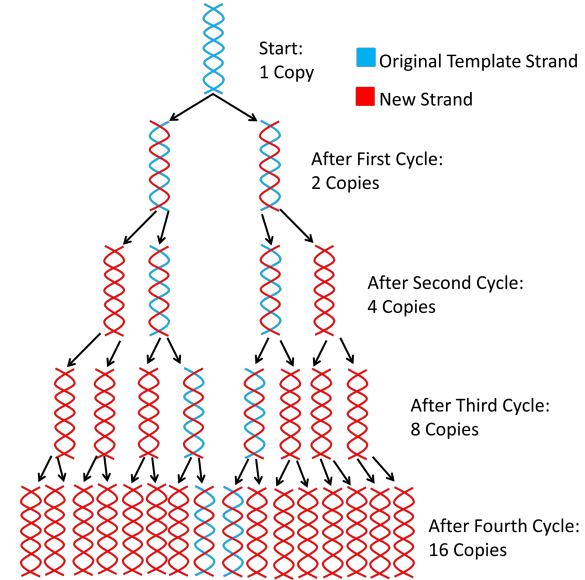
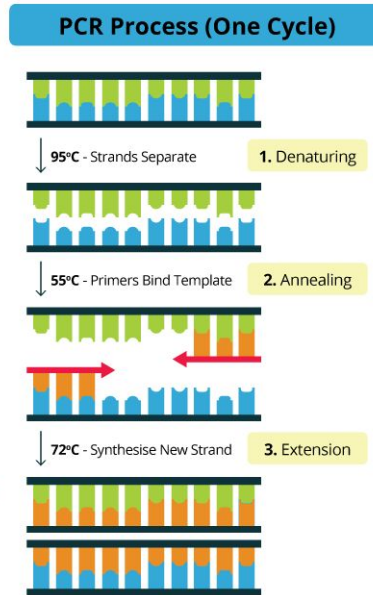
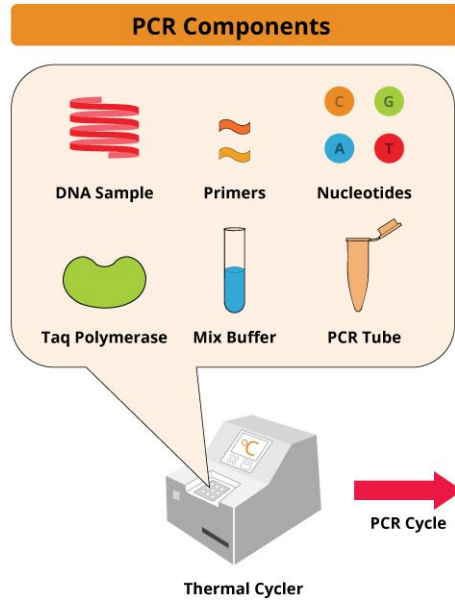
5'ggacgcaagc-----aggacattga 3'
3'cctgcgttcgac----- tcctgtaact 5'

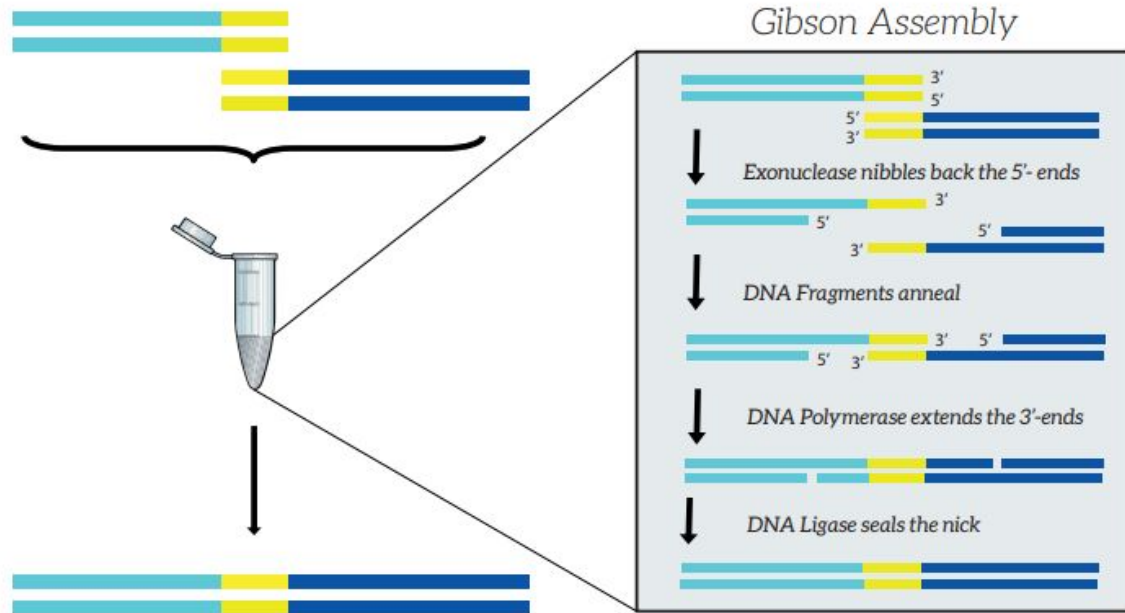
Desired sequence

5'gga**TC**caagc-----aggacattga 3'
3'cct**AG**gttcgac----- tcctgtaact 5'

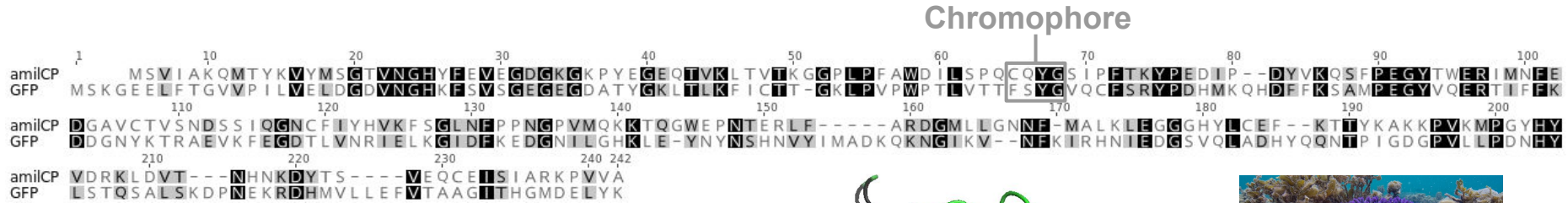


Copying DNA with Polymerase Chain Reaction





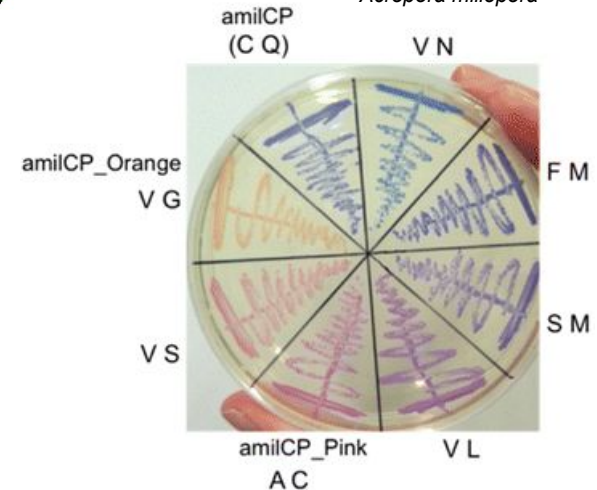
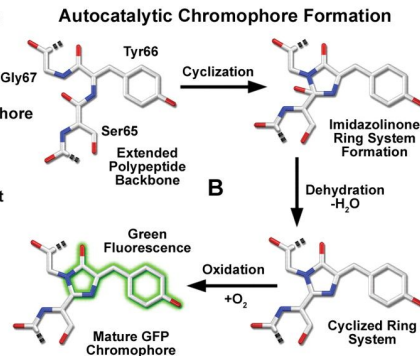
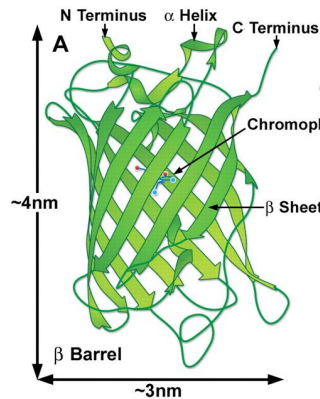
Creating the Chromophore Colorwheel



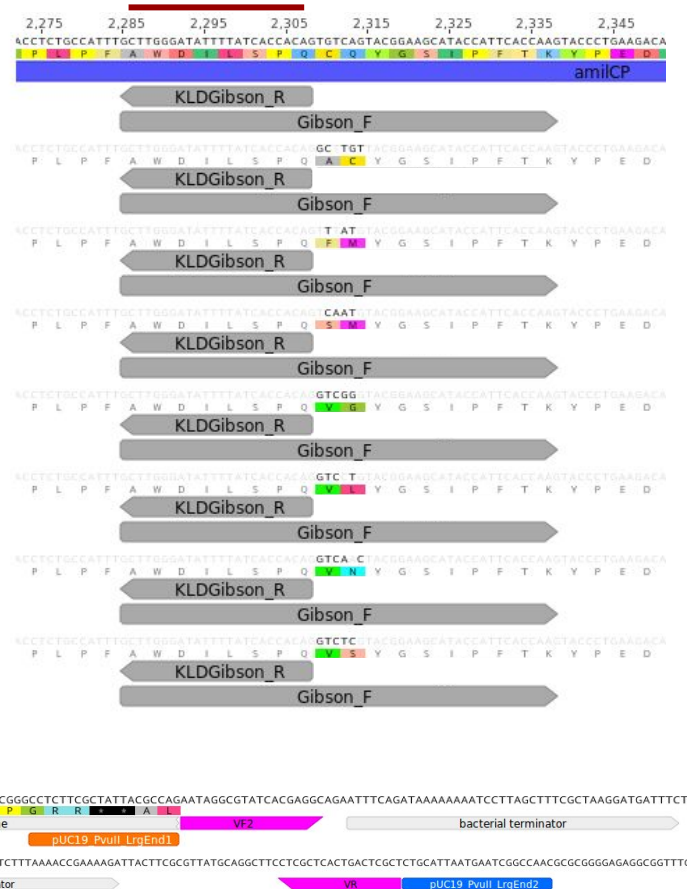
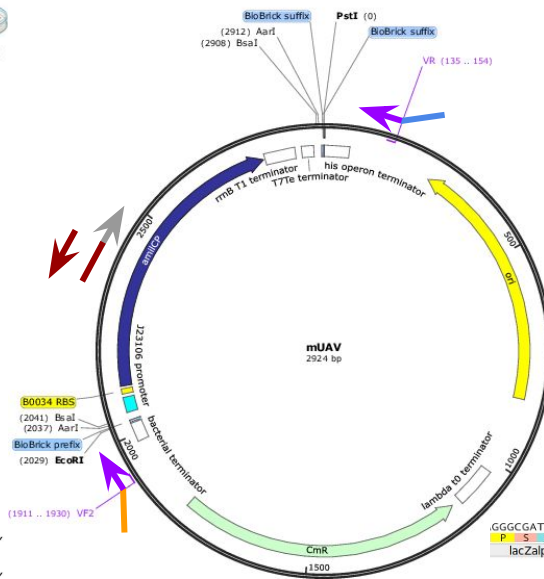
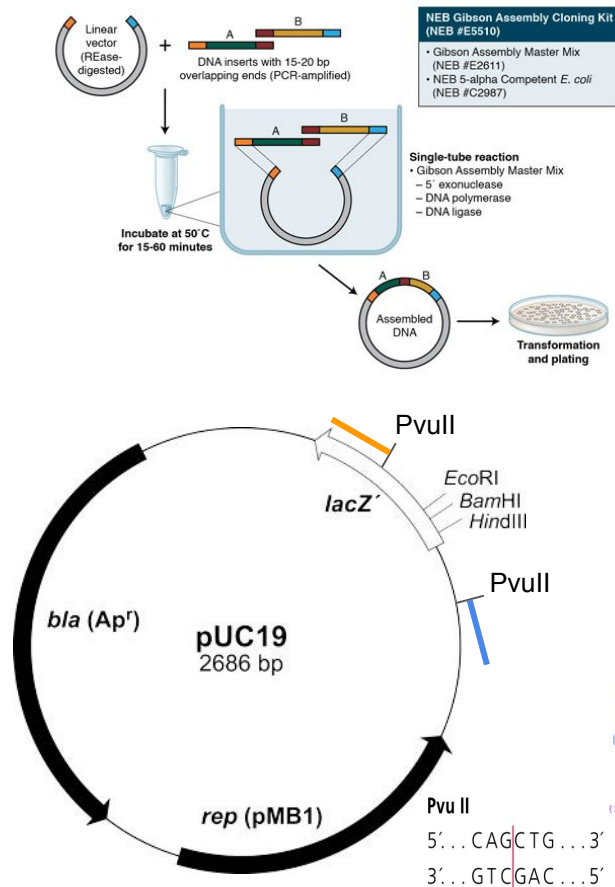
Conserved Similar Divergent



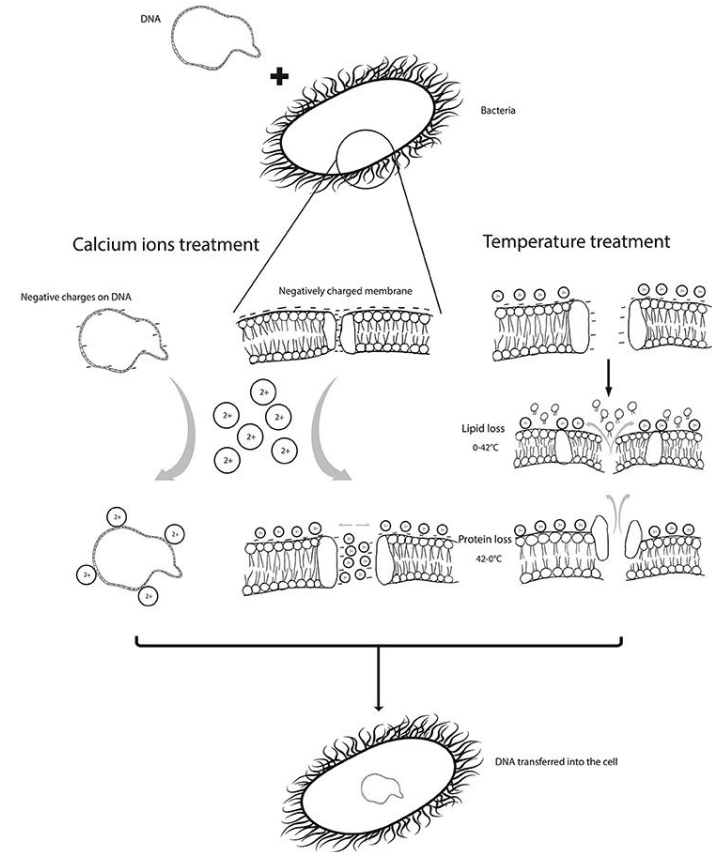
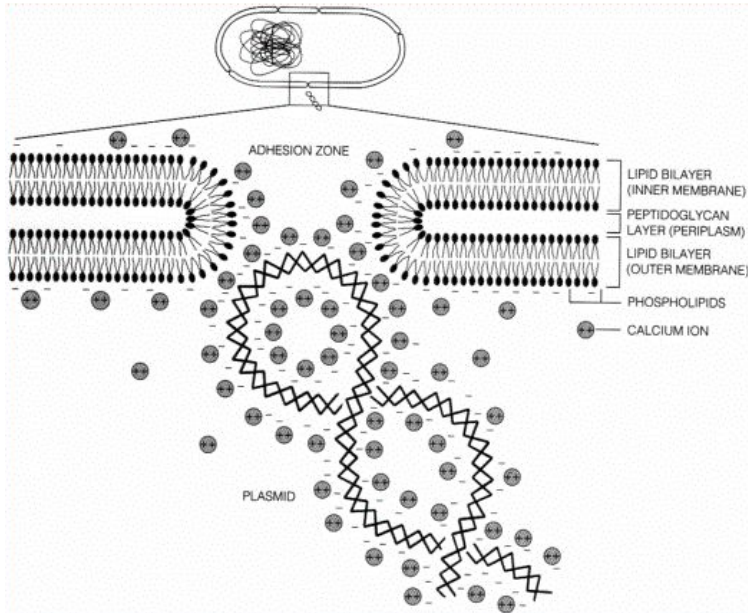
Acropora millepora



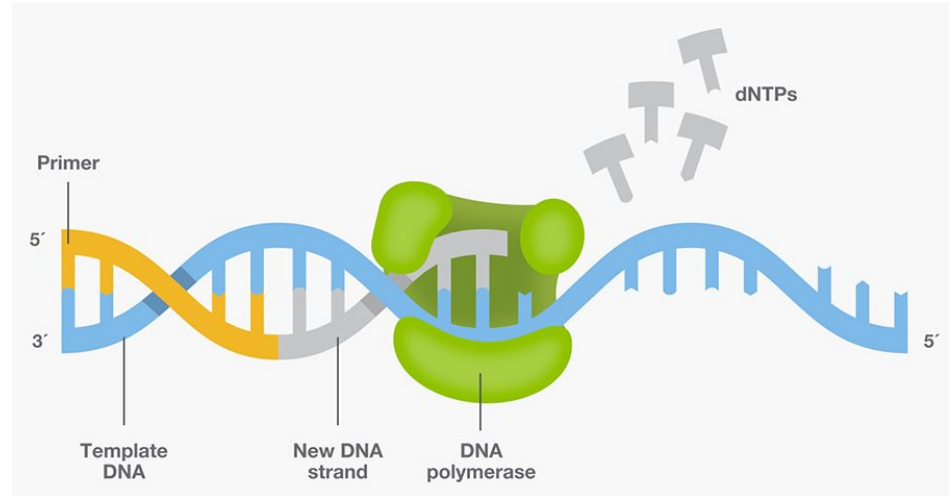
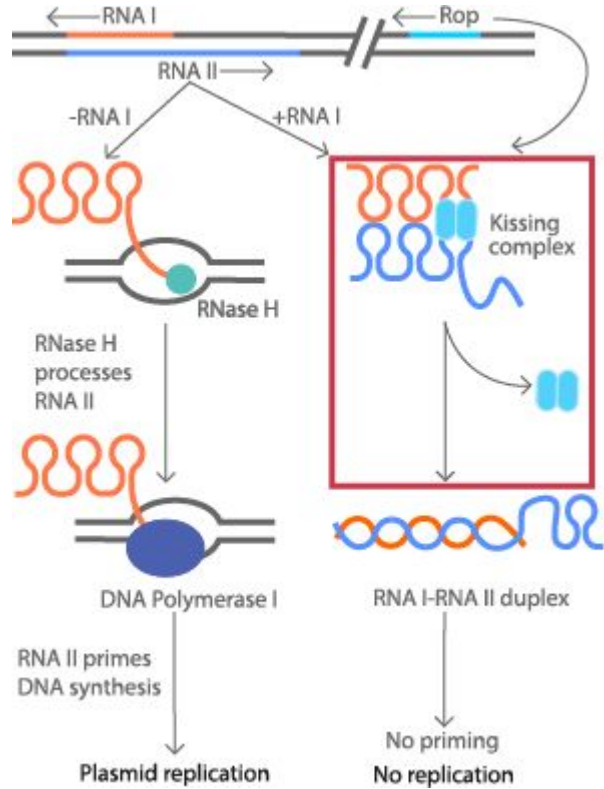
Gibson Assembly for Mutagenesis and Drug Selection Swap



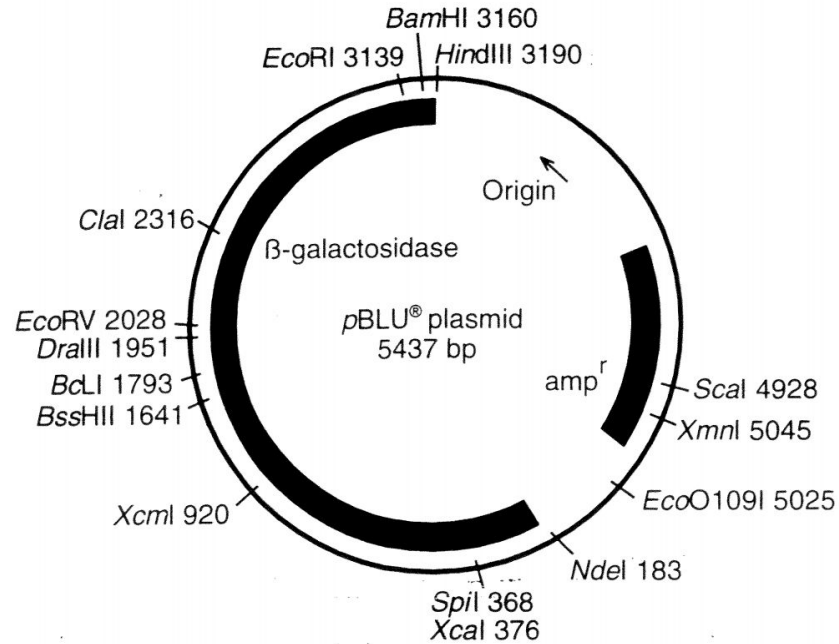
Plasmid Uptake by Calcium-Assisted Heat Shock



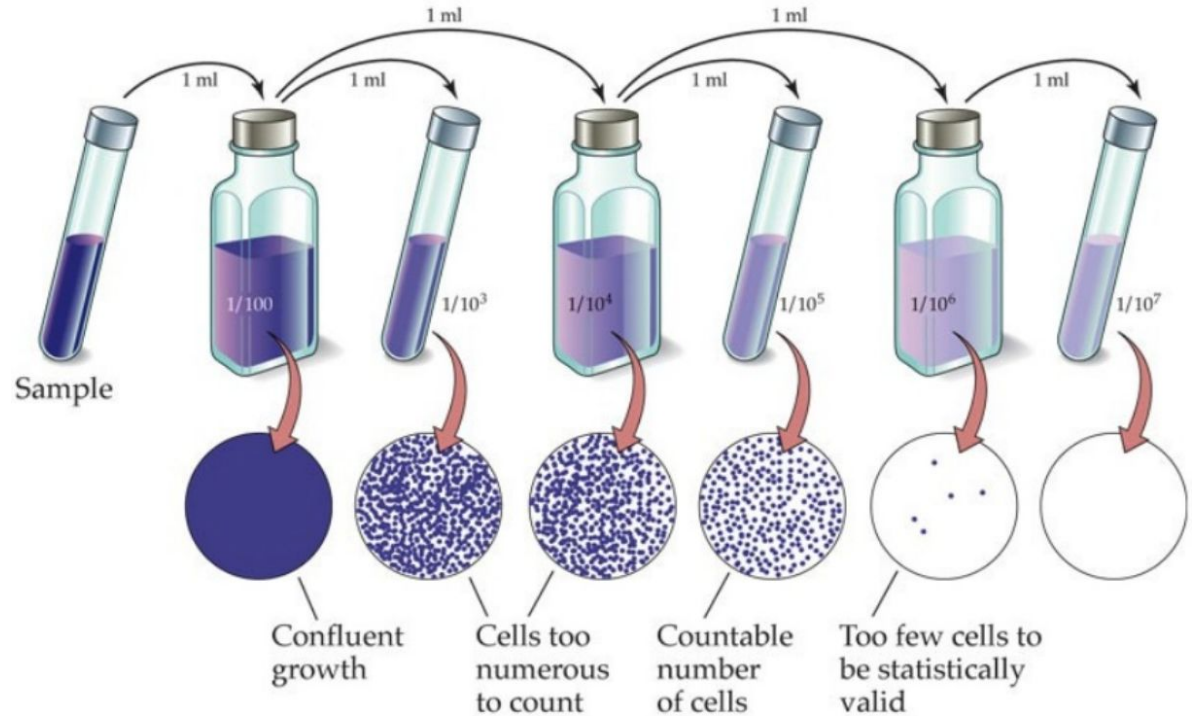
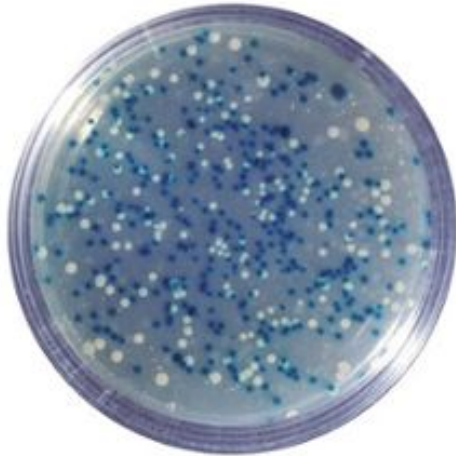
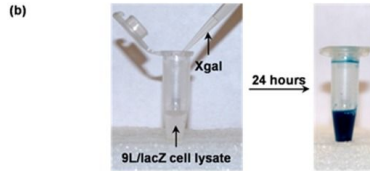
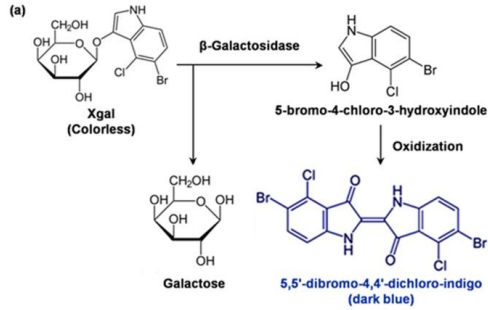
A Primer for This Week



DNA-Programming w/ Plasmids



Viability Count (Colony Forming Units, CFU) Serial Dilution

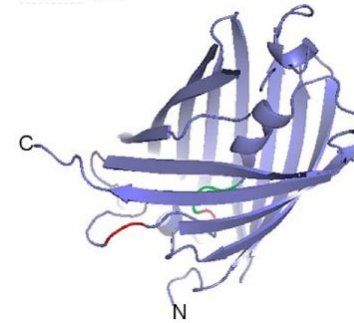


NextGenSynth Assignment:

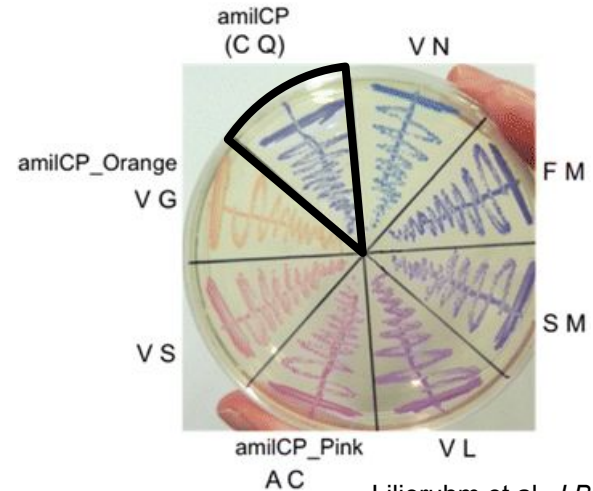
DNA-Programmed Coral Color Changes

Think like a molecule.
-George Church

Acropora millepora



Acropora millepora **ChromoProtein**

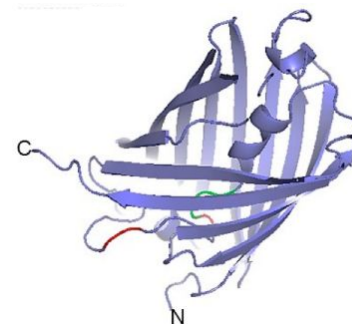
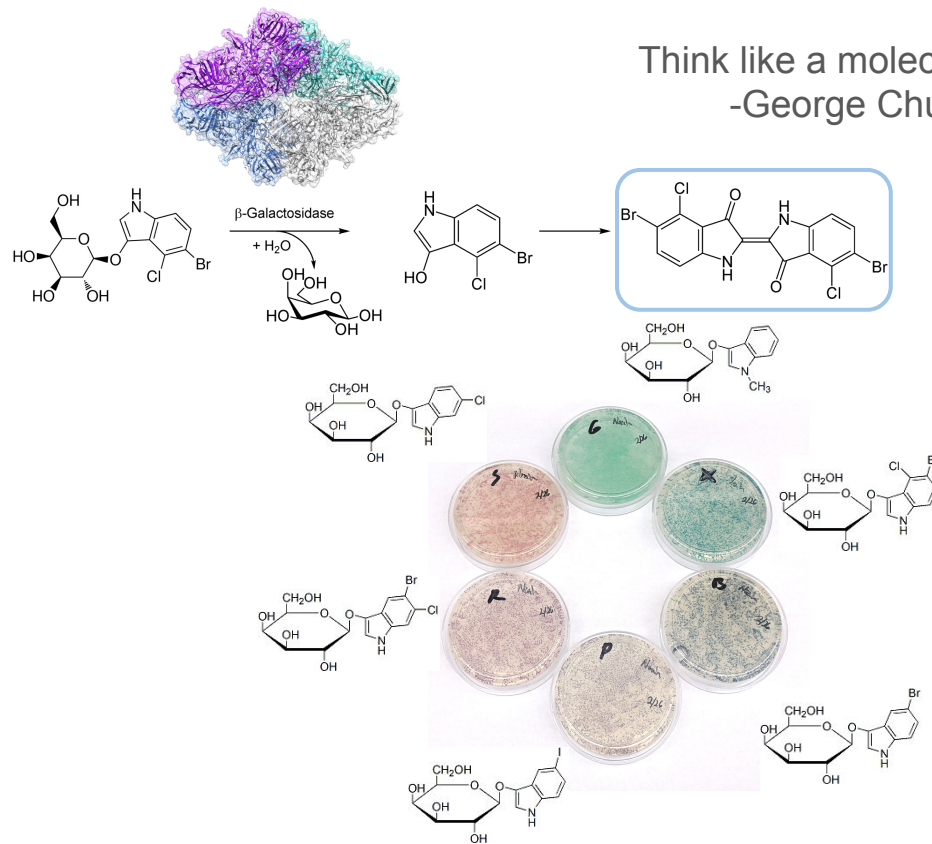


Liljeruhm et al, *J Biol Eng.* 2018

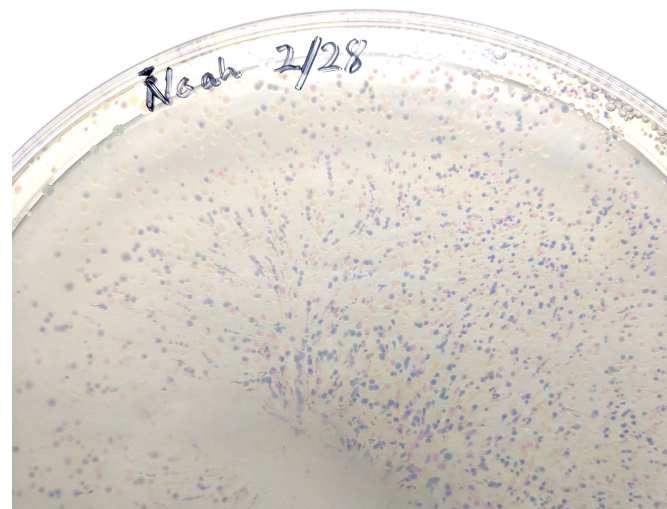
NextGenSynth Assignment:

DNA-Programmed Coral Color Changes

Think like a molecule.
-George Church



Acropora millepora ChromoProtein



Molecular Machinery: A Tour of the Protein Data Bank

Cells build many complex molecular machines that perform the biological jobs needed for life. Some of these machines are molecular scissors that cut food into digestible pieces. Others then use these pieces to build new molecules when cells grow or tissues need to be repaired. Some molecular machines form sturdy beams that support cells, and others are motors that use energy to crawl along these beams. Some recognize attackers and mobilize defenses against infection.

Researchers around the world are studying these molecules at the atomic level. These 3D structures are freely available at the Protein Data Bank (PDB), the central storehouse of biomolecular structures. A few examples from the ~100,000 structures held in the PDB are shown here at a magnification of about 3,500,000 times, with each atom represented as a small sphere. The enormous range of molecular sizes is illustrated here, from the water molecule (H₂O) with only three atoms (shown at the left) to the ribosomal subunits with hundreds of thousands of atoms.

Digestive Enzymes: breaking food into small nutrient molecules

1. Amylase 1a01
2. Pepsin 1pep
3. Trypsin 2trp
4. Lysosome 1l1t
5. Nucleosome 1nuc
6. Carboxypeptidase 3cpa
7. Ribonuclease 1rnc

Blood Plasma Proteins: transporting nutrients and defending against injury

8. Factor X 1xka, 1xkd
9. Thrombin 1tbr
10. Fibrin 1fm1, 2fml
11. Serum Albumin 1alb
12. Serum Albumin 1alb

Viruses and Antibodies: engaging in constant battle in the bloodstream

13. Antibody 1igt
14. Rhinovirus 4rhv

Hormones: carrying molecular messages through blood

15. Glucagon 1gon
16. Insulin 2ins
17. Epidermal Growth Factor 1egf

Channels, Pumps and Receptors: getting back and forth across the membrane

18. Ion Protein 1p21
19. Beta-Adrenergic Receptor/G-Protein 1bcr
20. Acetylcholine Receptor 2acr
21. Epidermal Growth Factor Receptor 1egf
22. Rhodopsin 1rhd
23. Polypeptide 1ac2
24. Potassium Channel 1kt
25. Calcium Pump 1puc
26. Cyclooxygenase 1p9t

Photosynthesis: harvesting energy from the sun

27. Photosystem II 1ps2
28. Light Harvesting Complex 1lhc
29. Photosynthetic Reaction Center 1pcc

Enzymes: cutting and joining the molecules of life

30. Fatty Acid Synthase 2fat
31. Bacterial Ribosome 1bct
32. Carboxylate/Dehydrogenase 1vca
33. Green Fluorescent Protein 1gfp
34. Luciferase 2luc
35. Glutamine Synthetase 2gls
36. Alcohol Dehydrogenase 2adh
37. Nitrogenase 1n2t
38. Laccase 1lac
39. beta-Lactamase 4blm
40. Catalase 1cat
41. Thymidine Synthase 2thc
42. Tryptophan Synthase 1wry
43. Aspartate Carboxyltransferase 4at

Storage: containing nutrients for future consumption

36. Ferritin 1ftr

Energy Production: powering the processes of the cell

39. Cytochrome c Oxidase (Complex IV) 1coo
40. Cytochrome c 1cyc
41. Cytochrome b1 (Complex III) 1b3g
42. Succinate Dehydrogenase (Complex II) 1suh
43. NADH Dehydrogenase (Complex I) 1nhd
44. ATP Synthase 1atp, 1atf, 1atp, 2atf
45. Myoglobin 1mbd
46. Hemoglobin 4hbh

Infrastructure: supporting and moving cells

63. Actin 1m9q
64. Myosin 1myo
65. Microtubule 1tub
66. Collagen 1col
- (see left)

Protein Synthesis: building new molecular machines

67. Transfer RNA 4trna
68. Valyl-tRNA Synthetase 1gas
69. Threonyl-tRNA Synthetase 1tgs
70. Glutamyl-tRNA Synthetase 1gls
71. Isoleucyl-tRNA Synthetase 1ily
72. Phenylalanyl-tRNA Synthetase 1pfs
73. Asparaginyl-tRNA Synthetase 1asn
74. Ribosome 1pke, 1p2
75. Elongation Factor Tu/tRNA 1te
76. Elongation Factor C 1efc
77. Elongation Factor Ts and Tu 1etu
78. Peptidyl 1p
79. Chaperone GroEL/ES 1am
80. Protein chaperone 1puc
81. Heat Shock Protein Hsp70 2ggs
82. Proteasome 4bap
83. Ubiquitin 1ubq

DNA: storing and reading genetic information

84. DNA 1dna
85. Restriction Endonuclease EcoRI 1eri
86. DNA Polymerase 1p1
87. Topoisomerase 1top
88. RNA Polymerase 1rpo
89. RNA Polymerase 1rpo
90. RNA Polymerase 1rpo
91. RNA Polymerase 1rpo
92. Single-stranded DNA-binding Protein 1ssb



Machines in the Microbial Factory

Escherichia coli

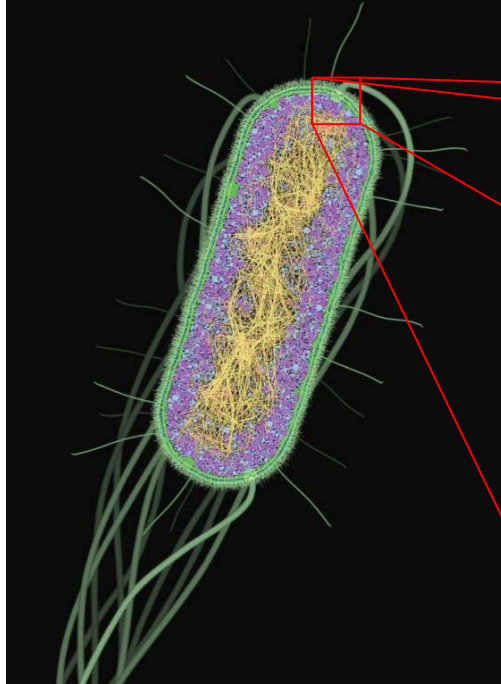


Image Credit: David Goodsell

Chromosome



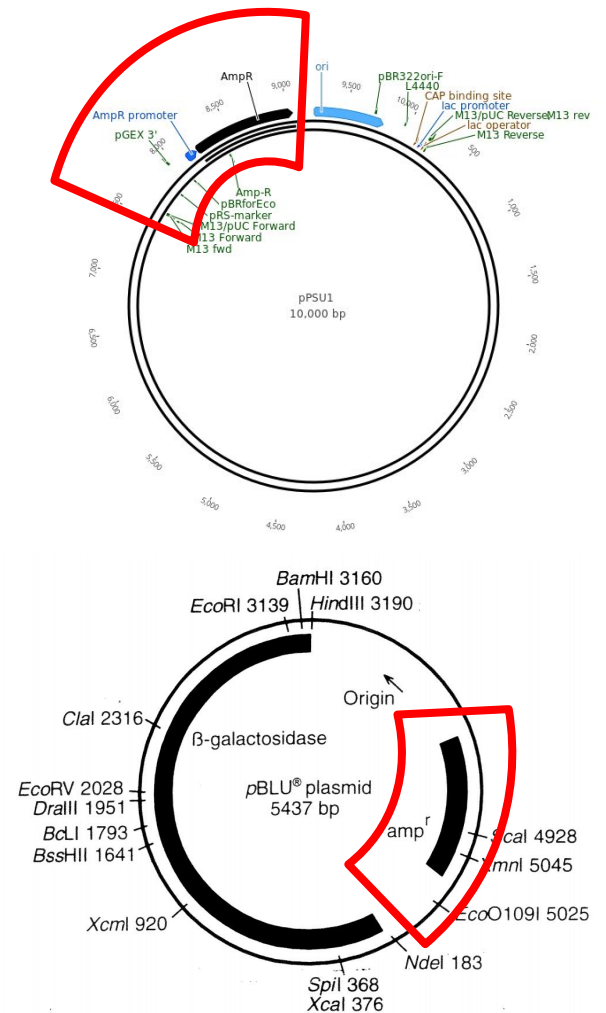
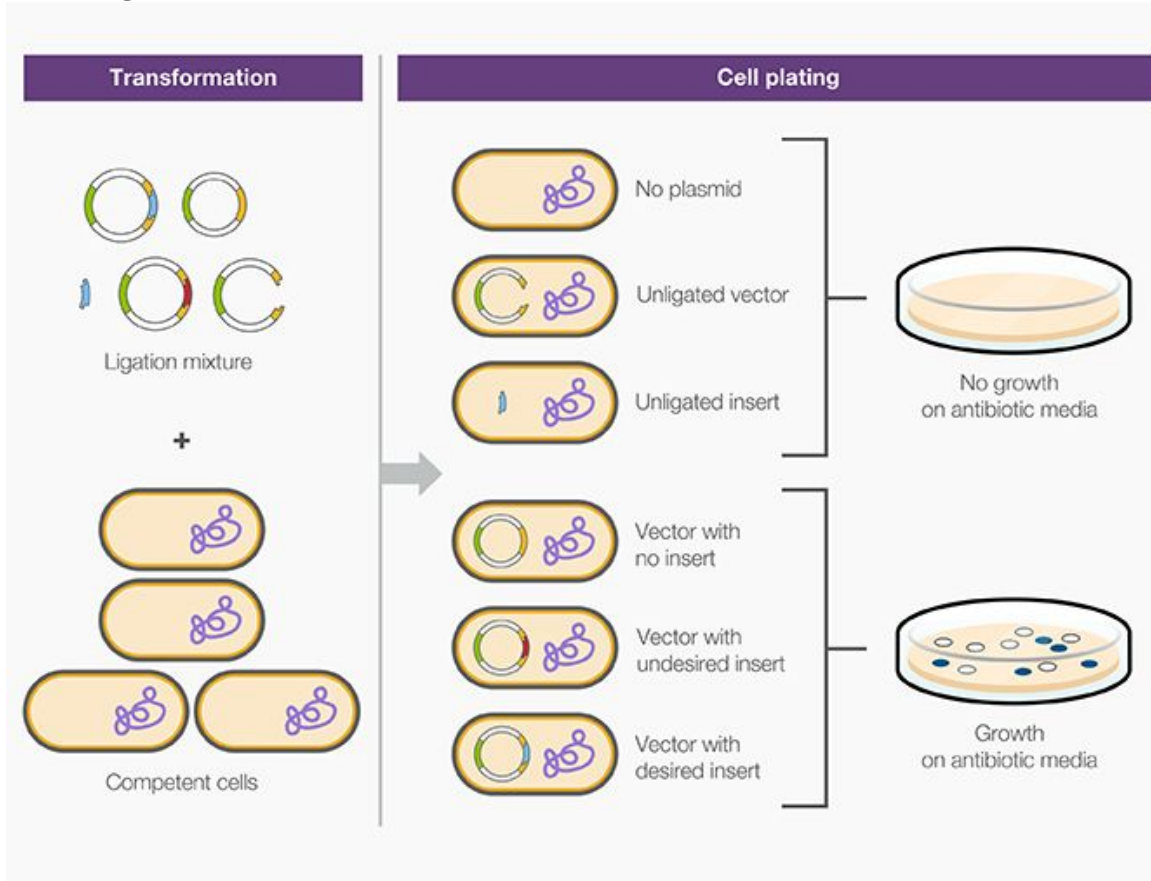
Flagellum

Inner Membrane

Cell Wall

Outer Membrane

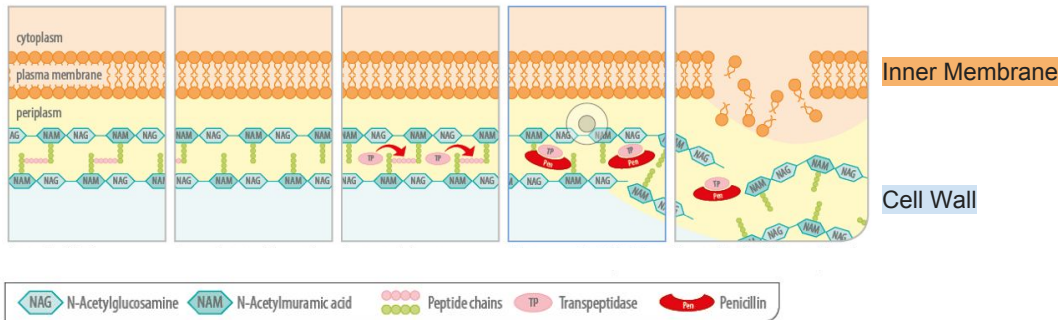
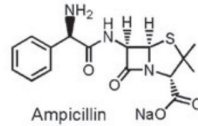
Drug Selection of Transformed *E coli* Cells



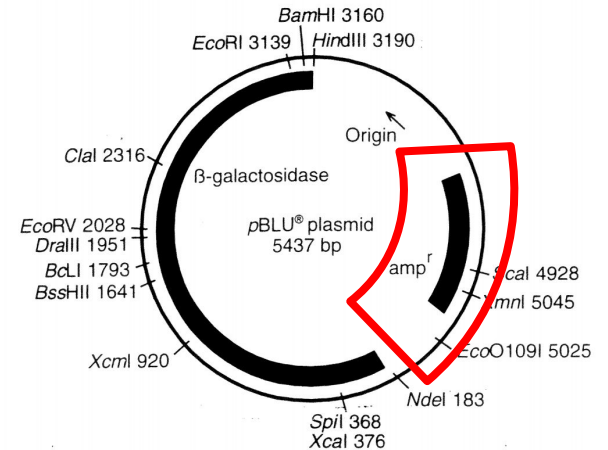
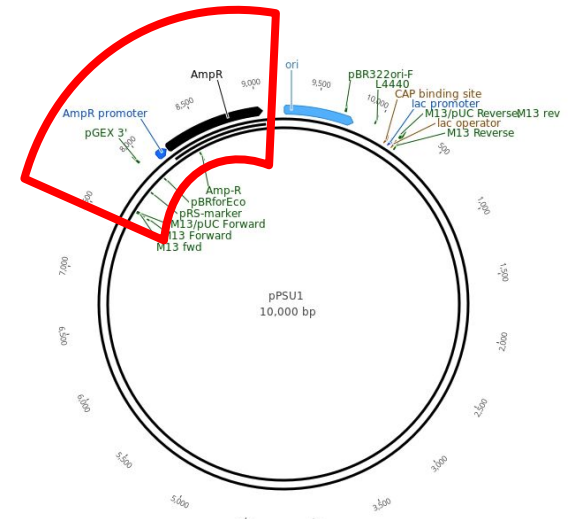
Drug Selection of Transformed *E. coli* Cells



Penicillium mold



*For simplicity, only two layers of peptidoglycans (NAM-NAG) are shown. Normally the cell wall consists of many layers.

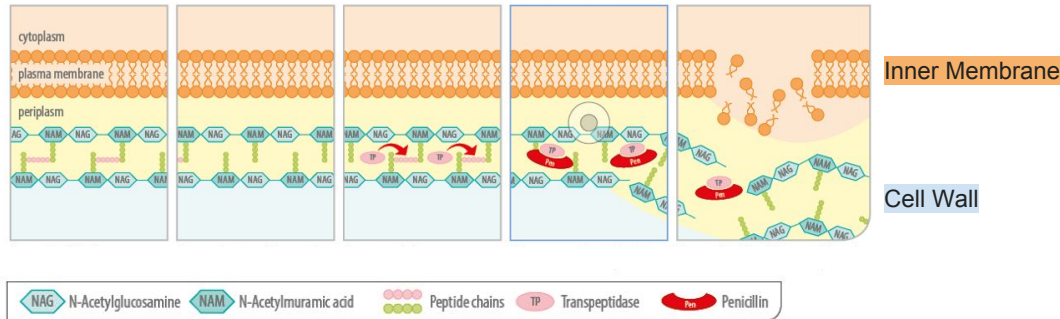
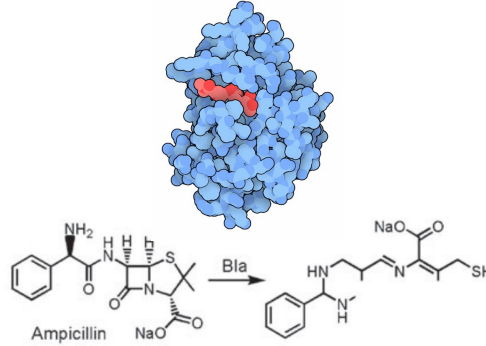


Drug Selection of Transformed *E coli* Cells

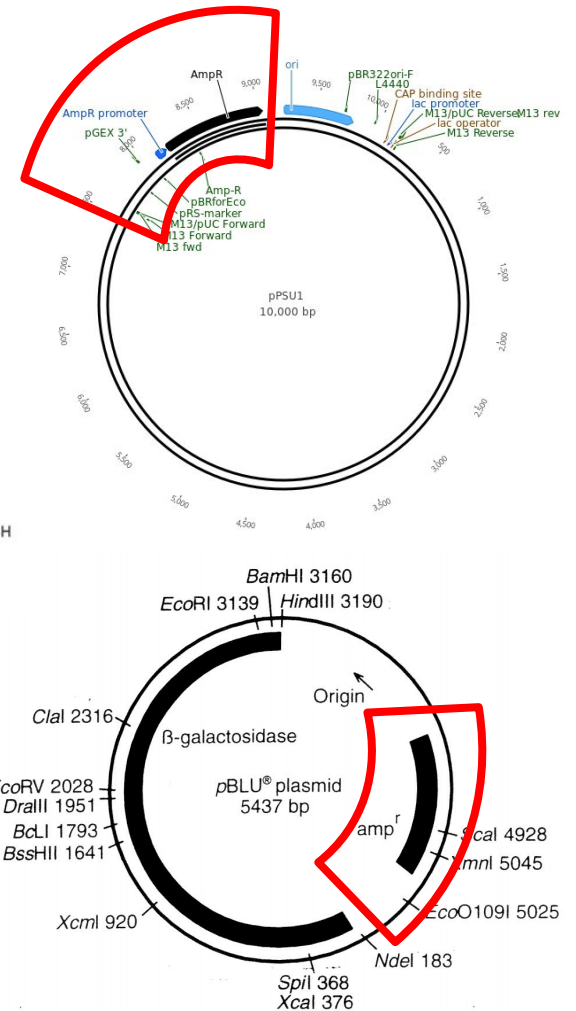


Penicillium mold

Beta-lactamase (Bla/amp^r)

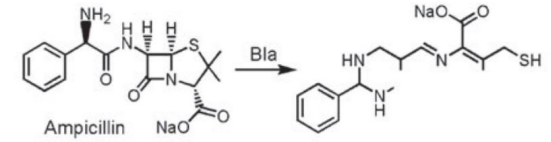
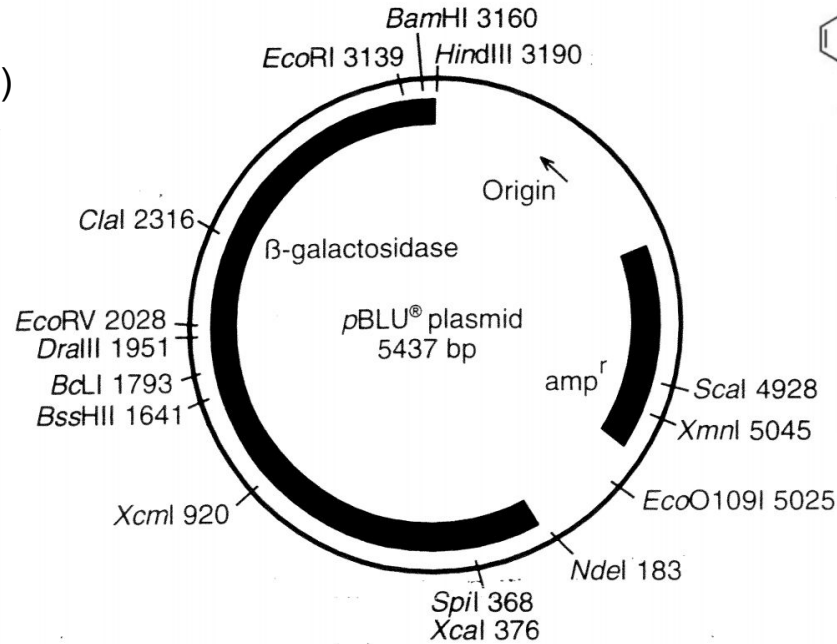
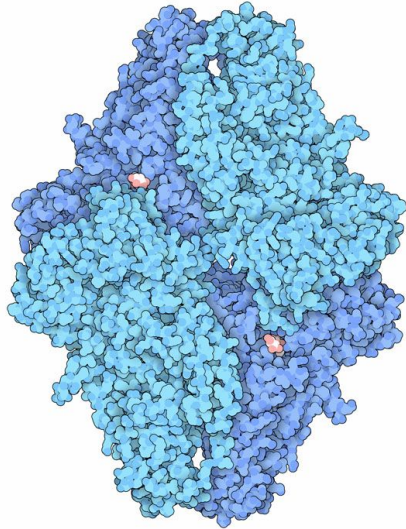


*For simplicity, only two layers of peptidoglycans (NAM-NAG) are shown. Normally the cell wall consists of many layers.

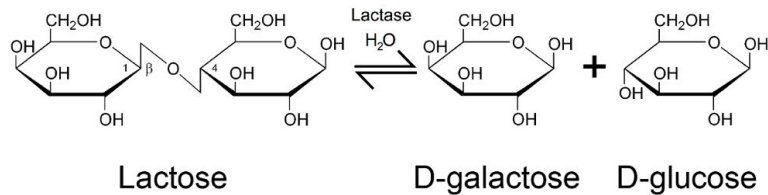
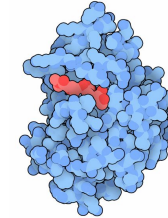


DNA-Programming w/ Plasmids

Beta-galactosidase (lacZ)

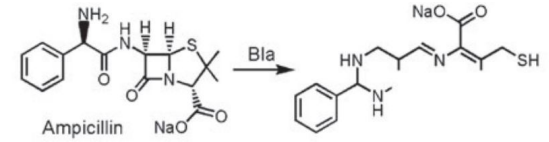
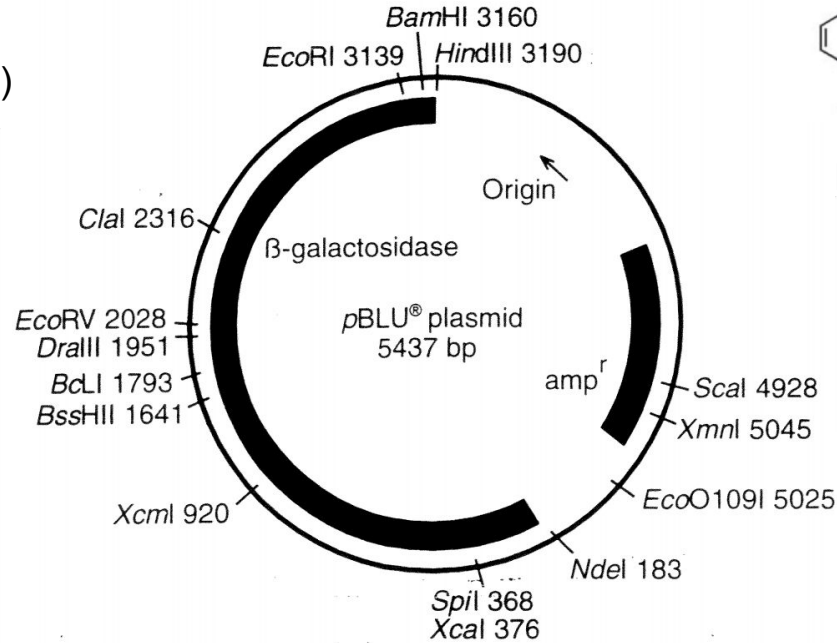
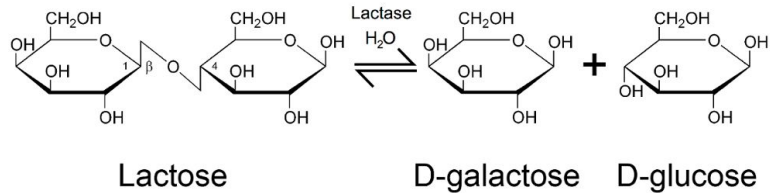
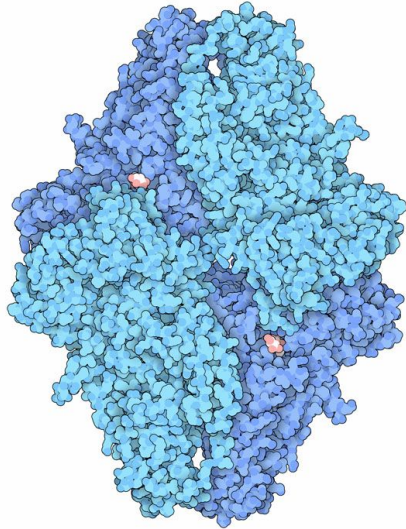


Beta-lactamase (Bla/amp^r)

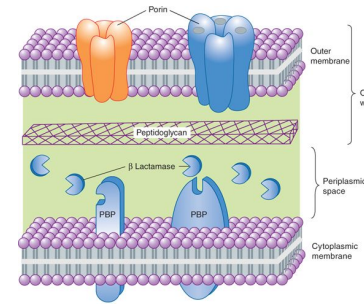
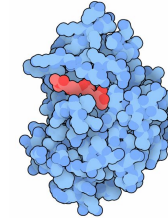


DNA-Programming w/ Plasmids

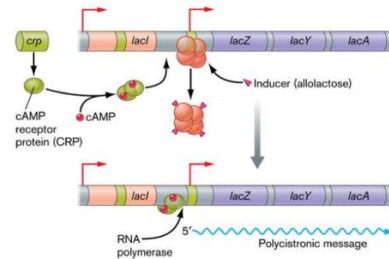
Beta-galactosidase (lacZ)



Beta-lactamase (Bla/amp^r)

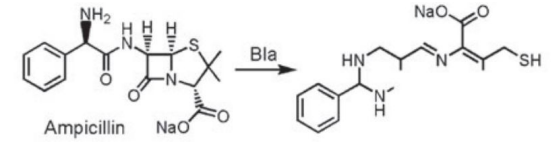
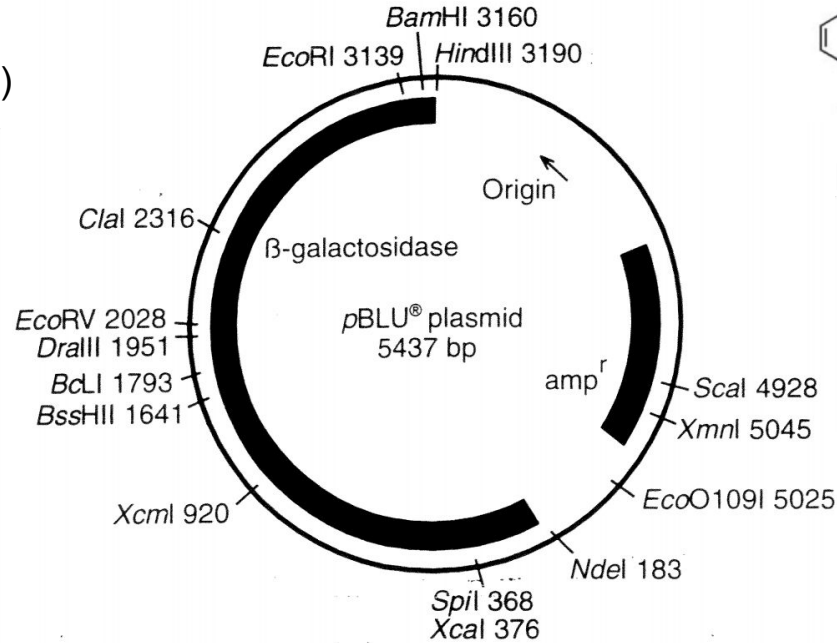
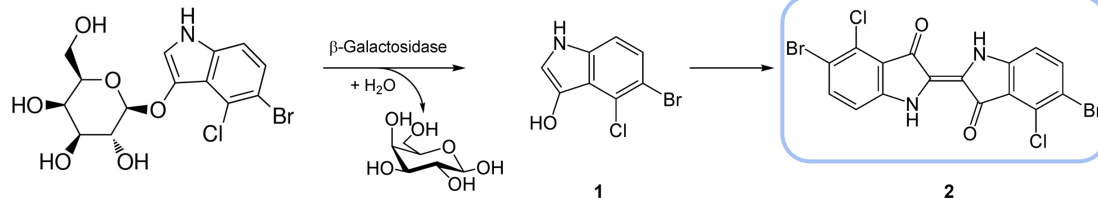
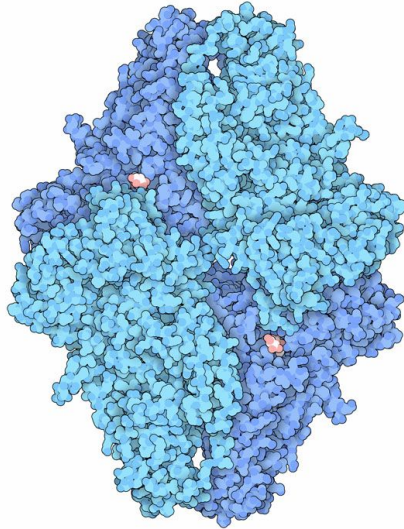


Source: Bertram G. Katzung, Anthony J. Trevor: Basic & Clinical Pharmacology, 13th Ed.
 www.accesspharmacy.com
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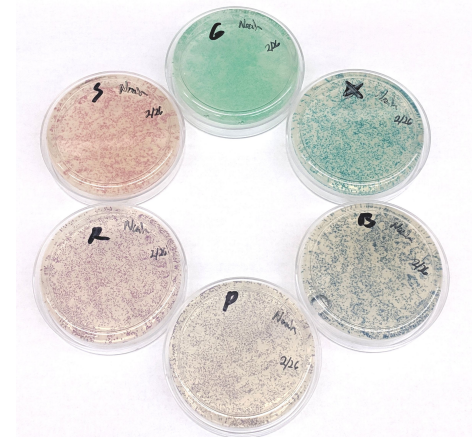
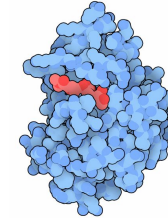


DNA-Programming w/ Plasmids

Beta-galactosidase (lacZ)

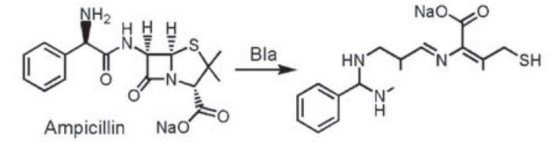
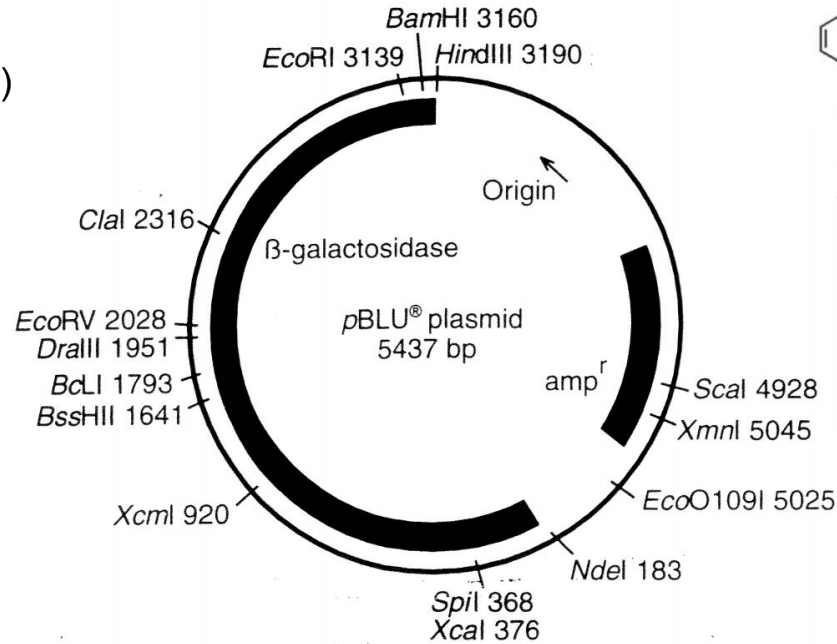
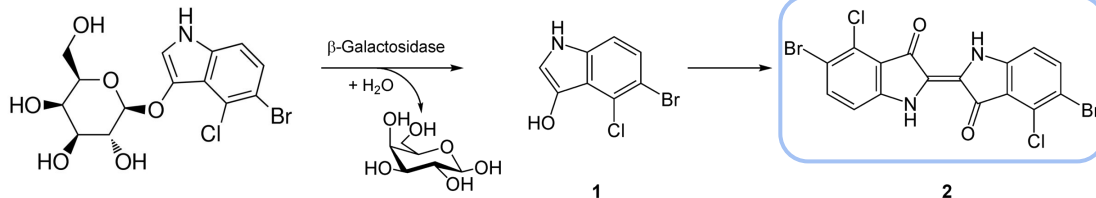
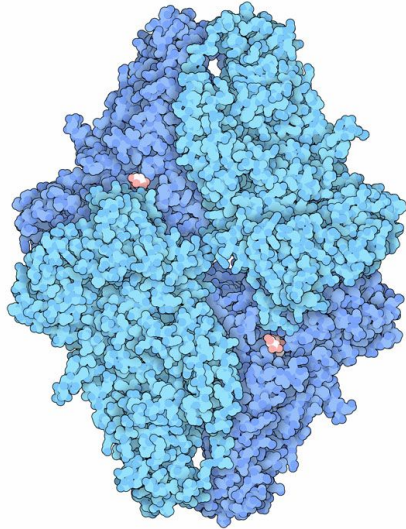


Beta-lactamase (Bla/amp^r)

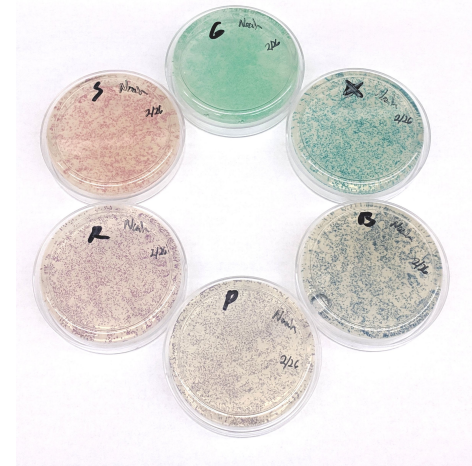
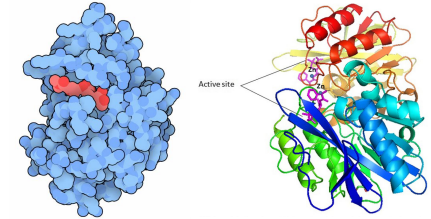


DNA-Programming w/ Plasmids

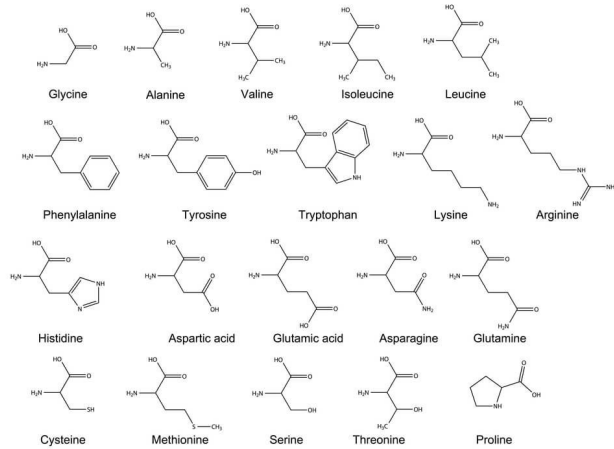
Beta-galactosidase (lacZ)



Beta-lactamase (Bla/amp^r)



Hierarchy of Protein Structure



(a) Primary

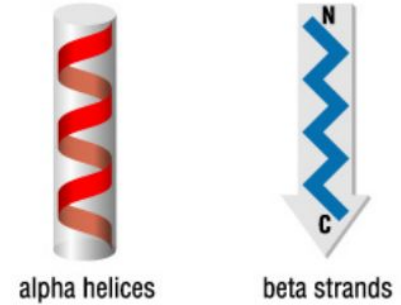


(c) Tertiary

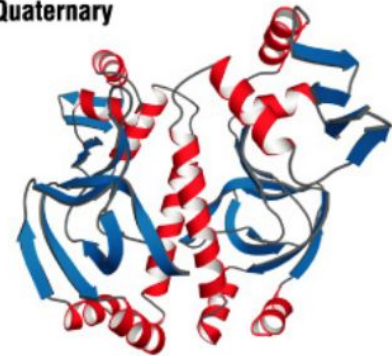


Cyclic-AMP Receptor Protein (CRP)

(b) Secondary

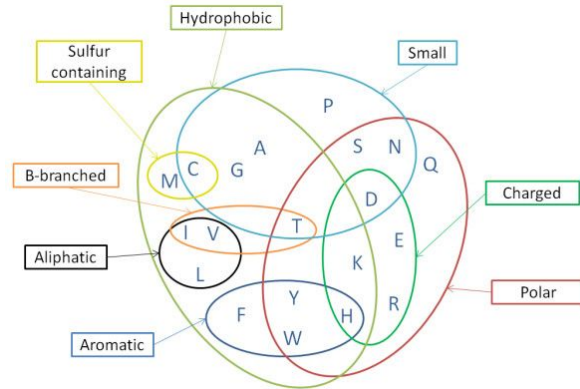


(d) Quaternary



Homodimer (CRP-CRP)

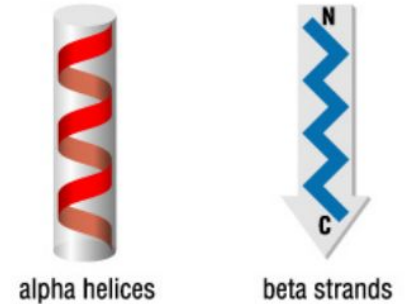
Hierarchy of Protein Structure



(a) Primary



(b) Secondary



(c) Tertiary



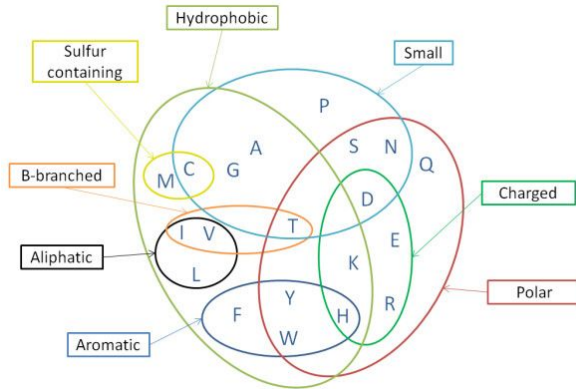
Cyclic-AMP Receptor Protein (CRP)

(d) Quaternary



Homodimer (CRP-CRP)

Hierarchy of Protein Structure

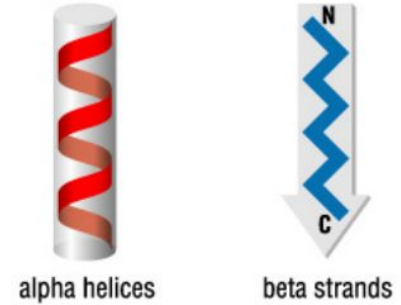


1: 686,323 (13%/13%)	2: 68,544 (6%/6%)	3: 31,472 (3%/3%)	4: 51,233 (3%/3%)	5: 70,713 (5%/5%)	6: 27,692 (4%/4%)
7: 7,474 (1%/0.7%)	8: 19,434 (2%/0.6%)	9: 26,462 (3%/0.5%)	10: 13,009 (3%/0.4%)	11: 384,921 (12%/0.4%)	12: 4,726 (0.9%/0.4%)
13: 6,180 (0.5%/0.4%)	14: 23,295 (3%/0.3%)	15: 16,468 (2%/0.3%)	16: 9,010 (1%/0.3%)	17: 16,575 (4%/0.3%)	18: 6,429 (1%/0.3%)
19: 16,981 (2%/0.3%)	20: 9,030 (0.5%/0.2%)	21: 23,099 (2%/0.2%)	22: 38,035 (4%/0.2%)	23: 20,517 (1%/0.2%)	24: 18,140 (3%/0.2%)
1: 686,323 (13%/13%)	2: 70,205 (6%/6%)	95: 18,778 (4%/0.06%)	608: 80 (0.04%/0.01%)	542: 303 (0.1%/0.01%)	
13: 6,586 (0.5%/0.4%)	10: 14,698 (1%/0.4%)	211: 263 (0.09%/0.03%)	84: 87 (0.06%/0.06%)	106: 290 (0.1%/0.05%)	
20: 9,030 (0.5%/0.2%)	66: 1,943 (0.4%/0.07%)	256: 1,006 (0.3%/0.02%)	119: 334 (0.2%/0.04%)	1019: 66 (0.04%/0.006%)	

(a) Primary



(b) Secondary

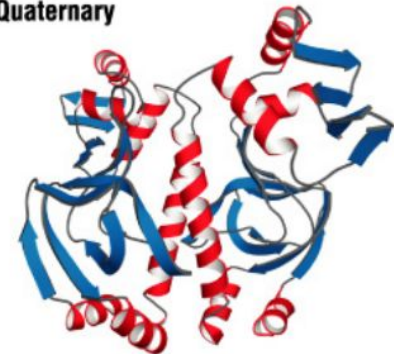


(c) Tertiary



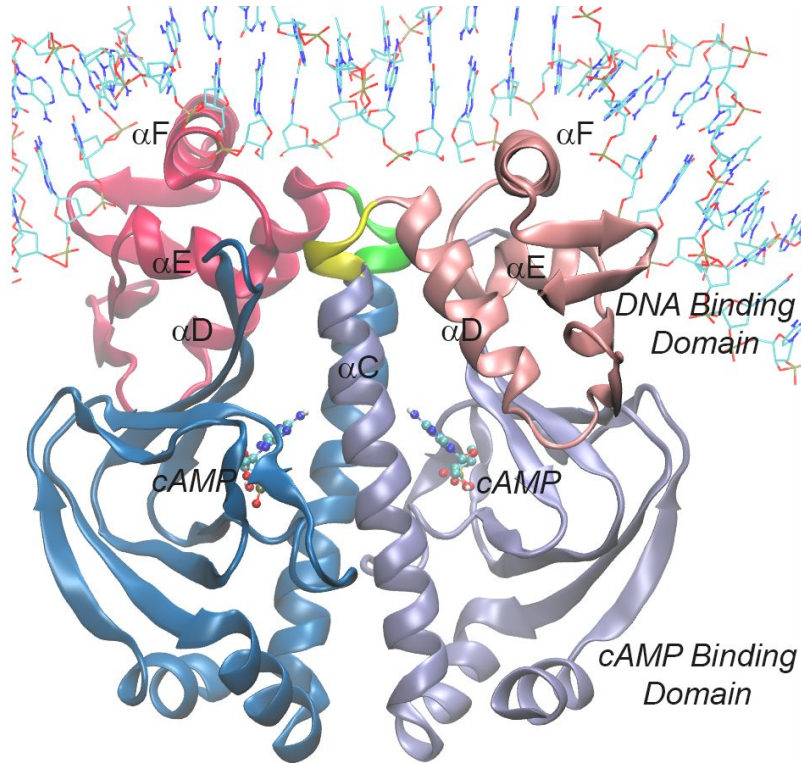
Cyclic-AMP Receptor Protein (CRP)

(d) Quaternary

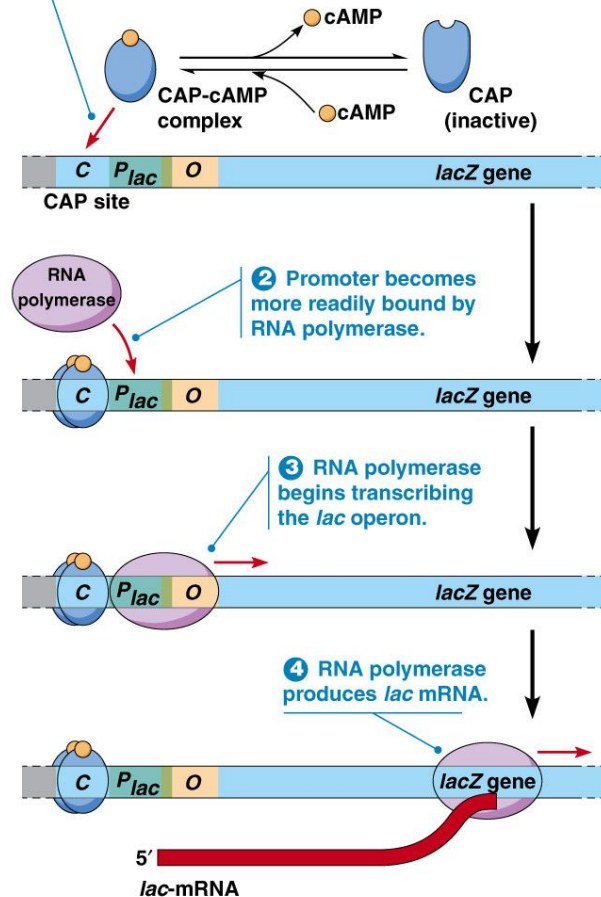


Homodimer (CRP-CRP)

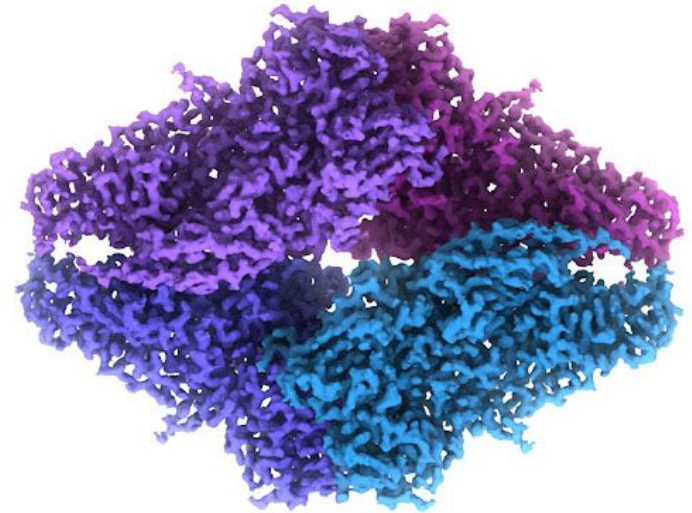
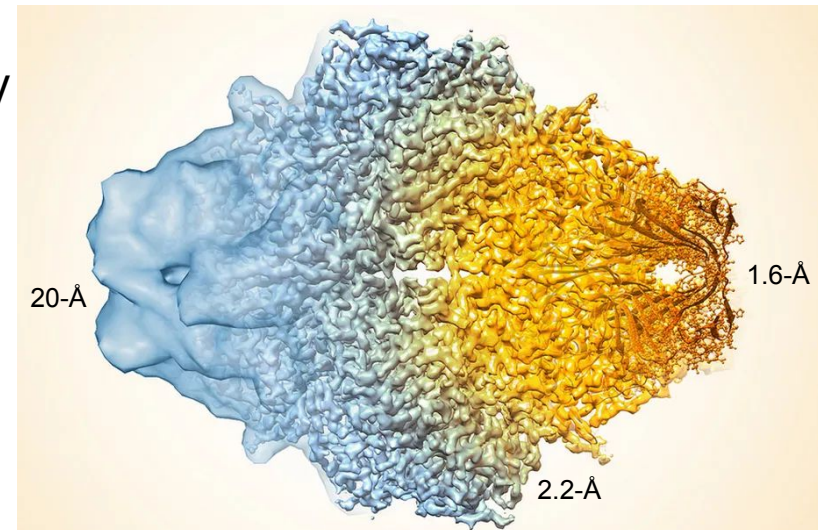
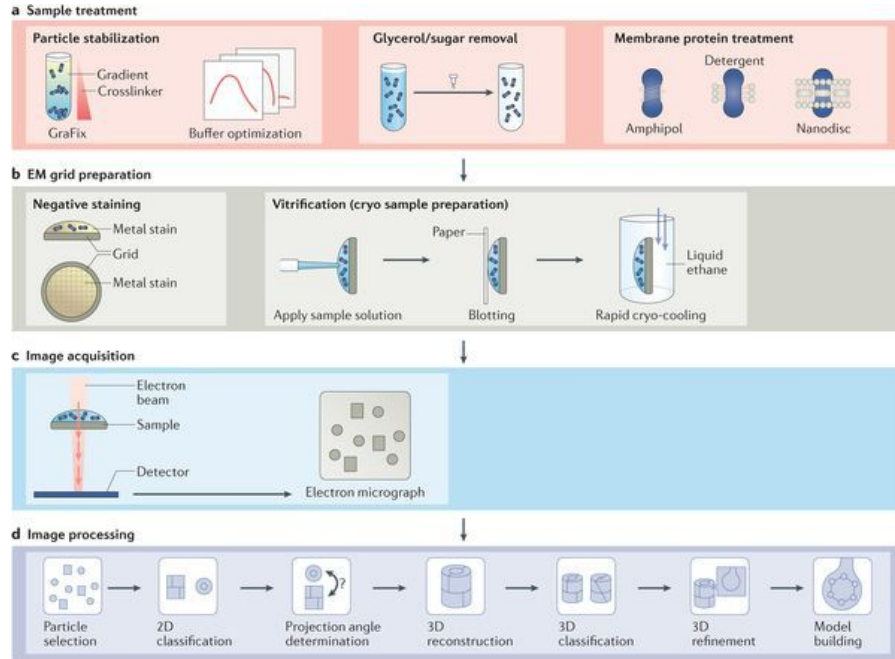
Modularity in Motifs and Domains



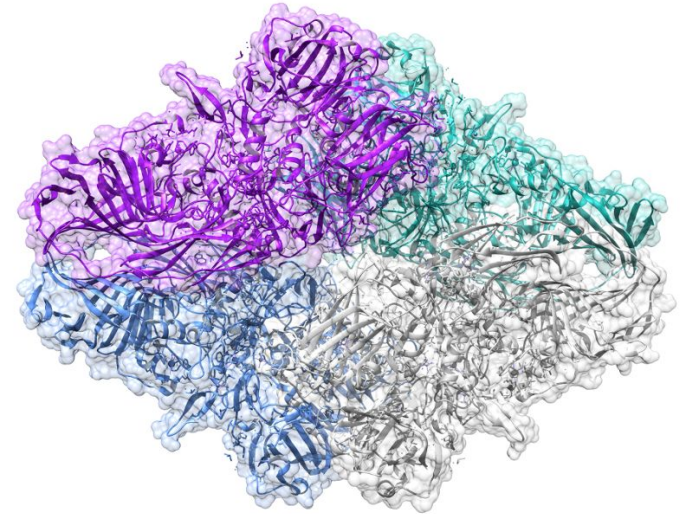
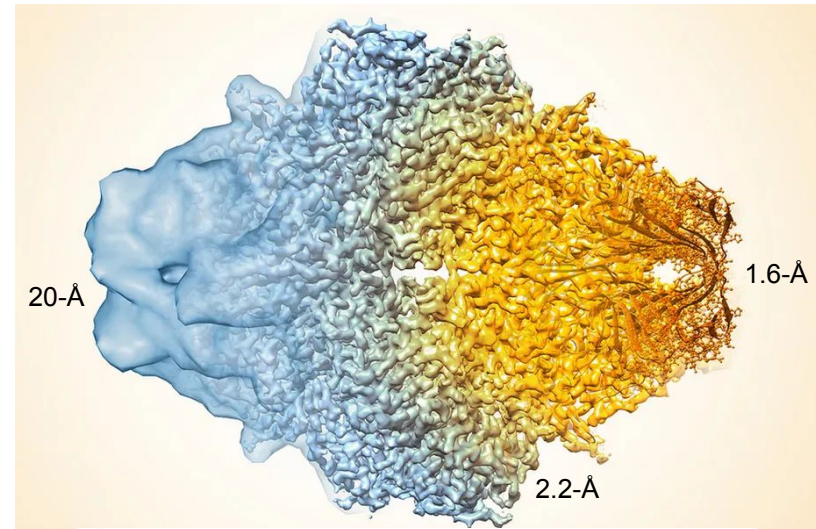
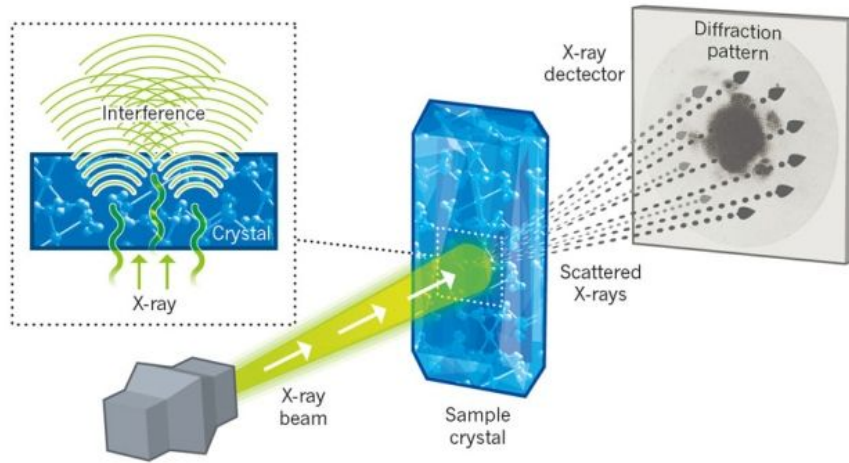
1 CAP-cAMP complexes bind to the CAP recognition site (C) near the promoter of the *lac* operon.



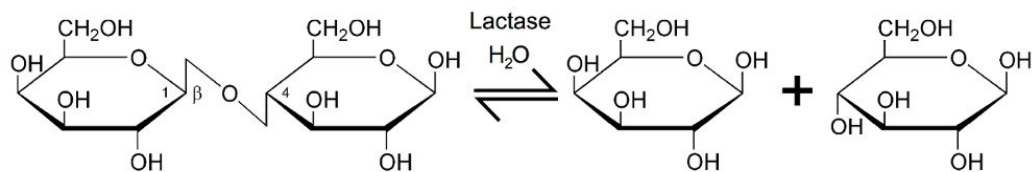
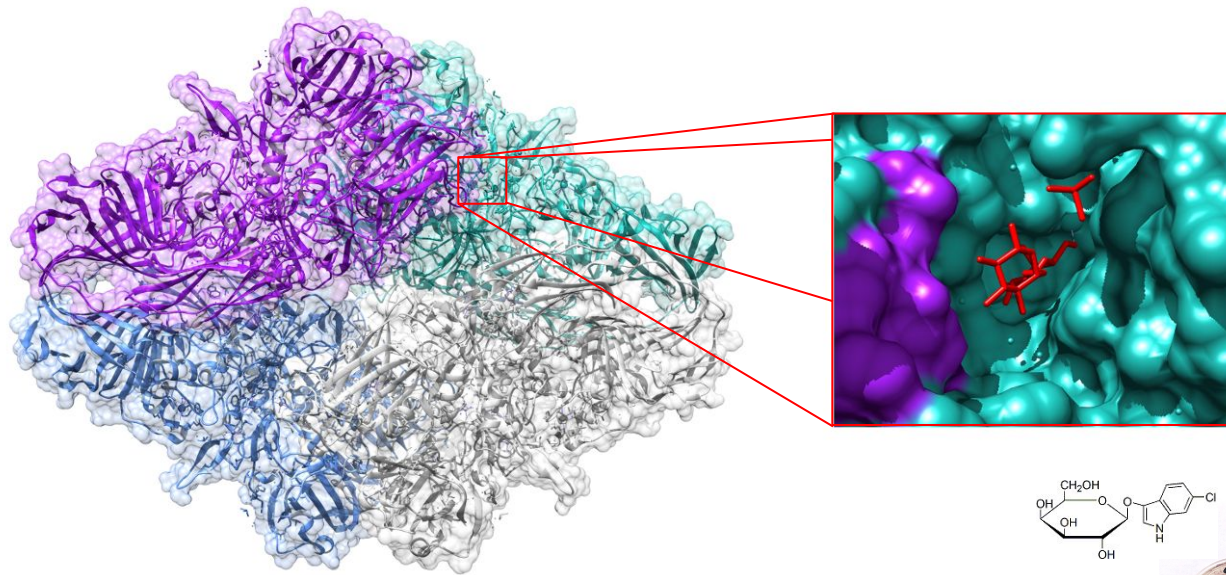
Solving for Structure: Cryo Electron Microscopy



Solving for Structure: X-Ray Crystallography



Color-Based Screens with Beta-galactosidase (lacZ)



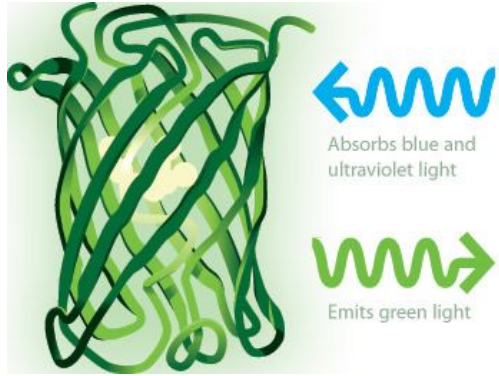
Lactose

D-galactose

D-glucose



Genetically-Encoded Green Fluorescence



Aequorea victoria

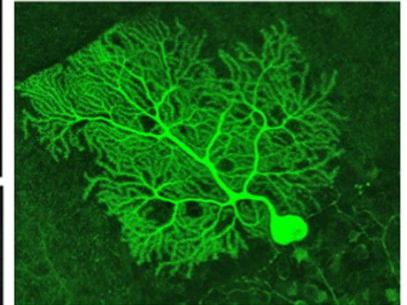
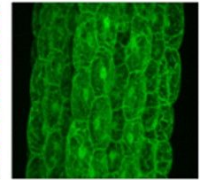
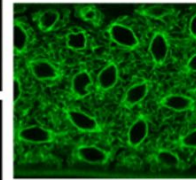
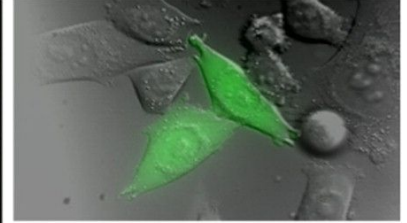
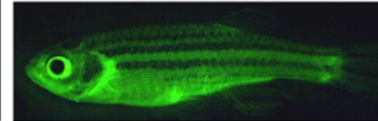
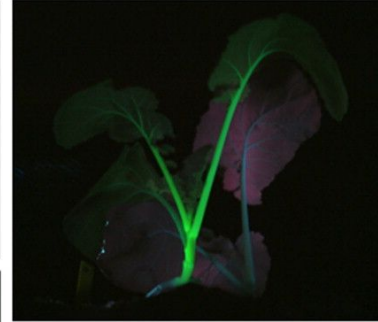
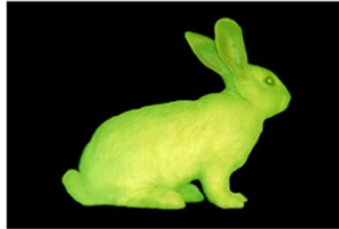
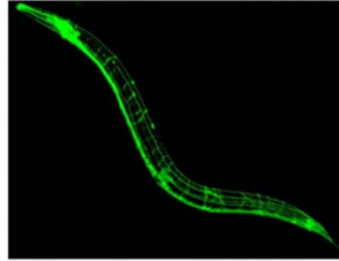
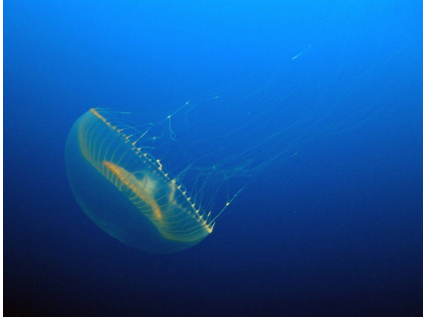


Image Credit: Roger Tsien Nobel Lecture

Chromophore Engineering

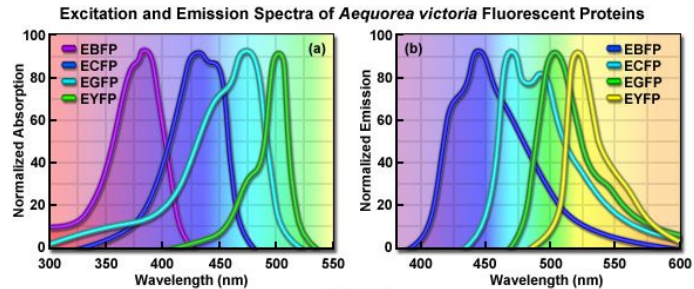
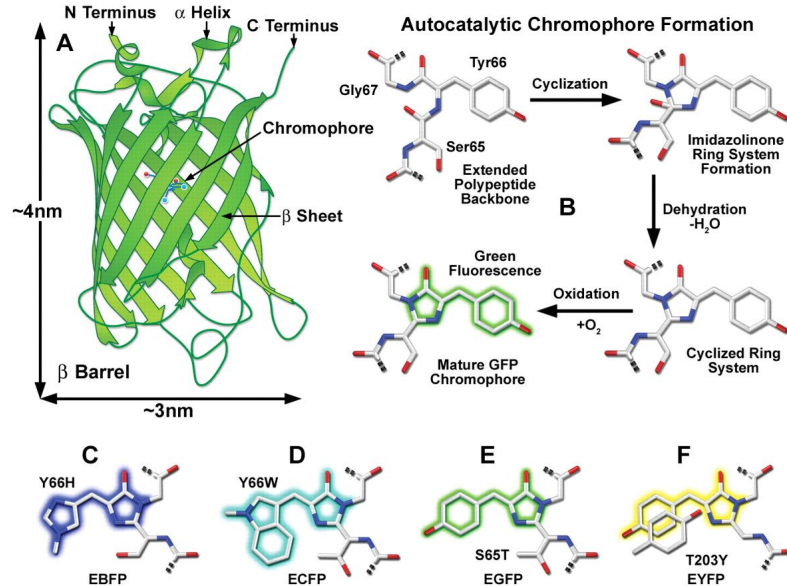
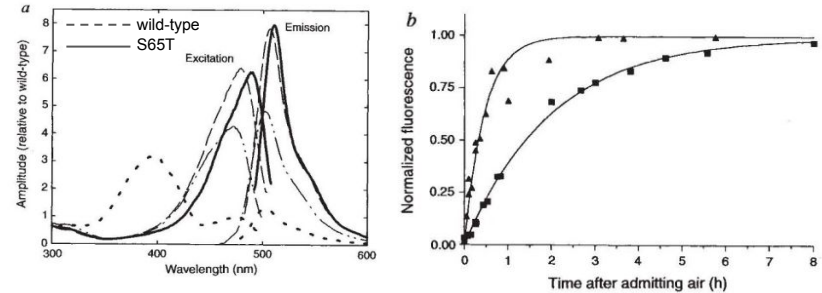
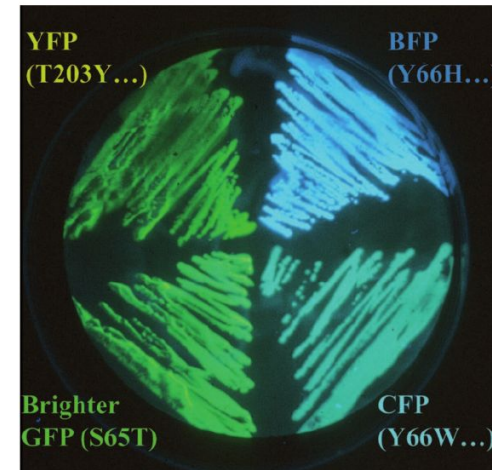


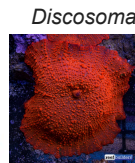
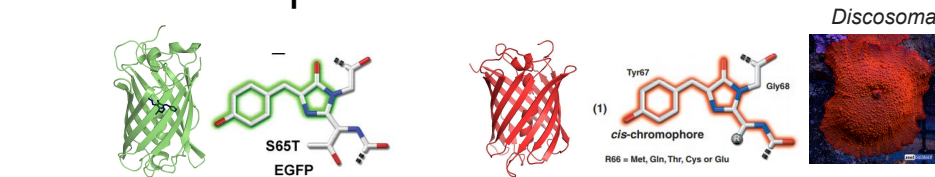
Figure 4



b, Rates of autooxidative fluorophore generation in wild-type (■) and S65T (▲) GFP, measured by development of fluorescence after admission of air to *E. coli* cultures anaerobically grown in GasPak pouches (Becton-Dickinson) for 3 days. Air was readmitted while transferring the cells to phosphate-buffered saline containing 8 mM $NaNO_3$ as a metabolic inhibitor. The time course of subsequent fluorescence development measured the final oxidation step in the protein's self-modification to generate its internal fluorophore⁸.



Fluorescent Spectrum & Structures



Arabidopsis thaliana

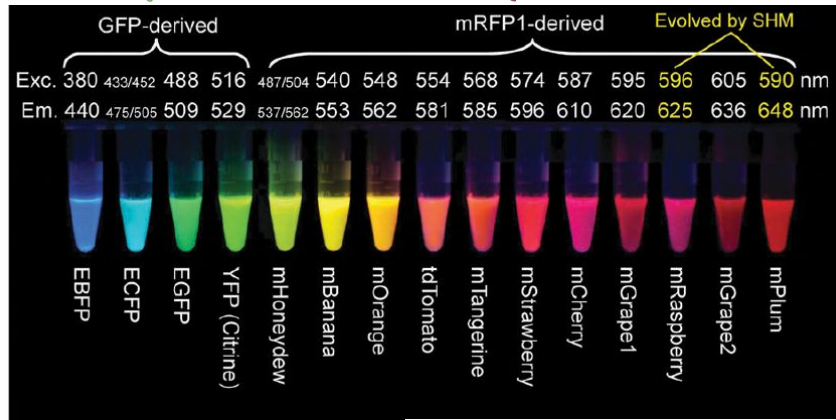
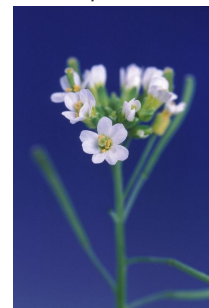
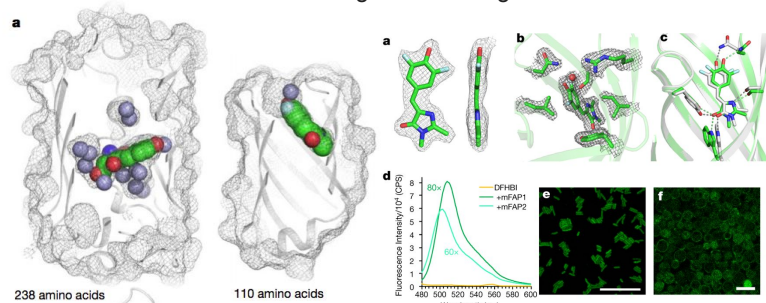
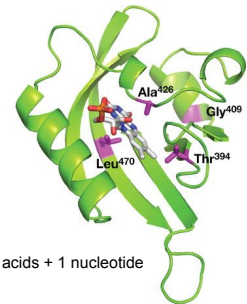
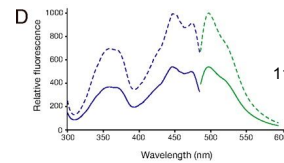
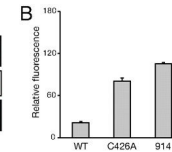
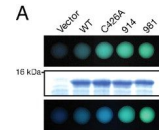


Image Credit: Roger Tsien Nobel Lecture



Dou et al, *Nature* 2018



110 amino acids + 1 nucleotide

Chapman et al, *PNAS* 2008

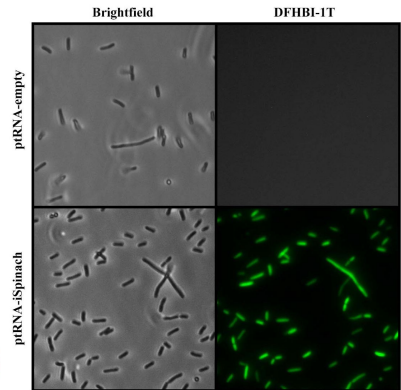
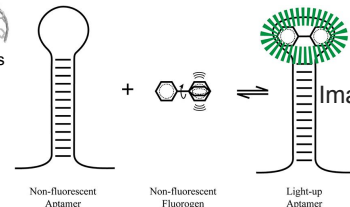
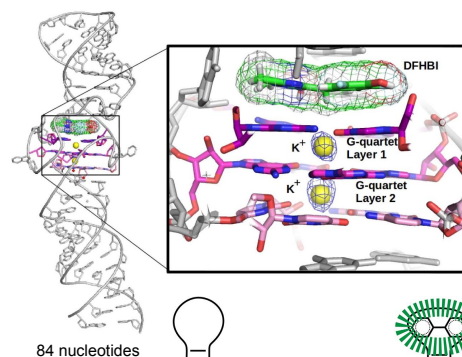
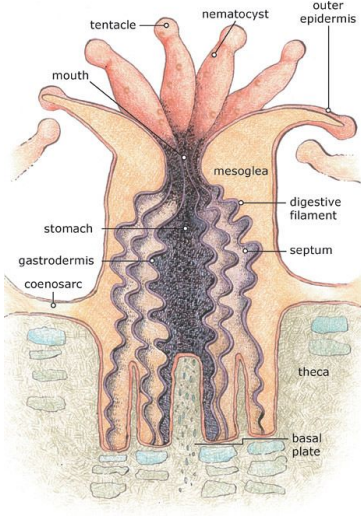


Image Credit: Roger Tsien Nobel Lecture

Protein Pigments



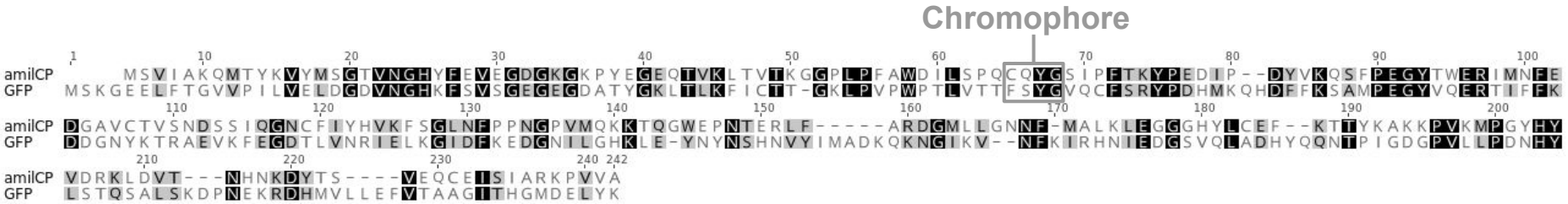
Acropora millepora

Protein Pigment Pellet Palette

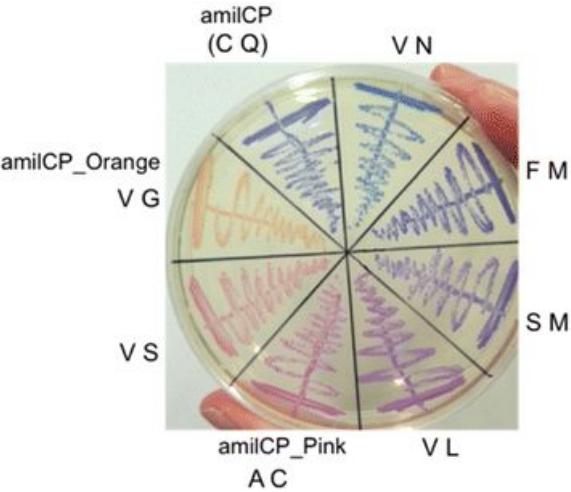
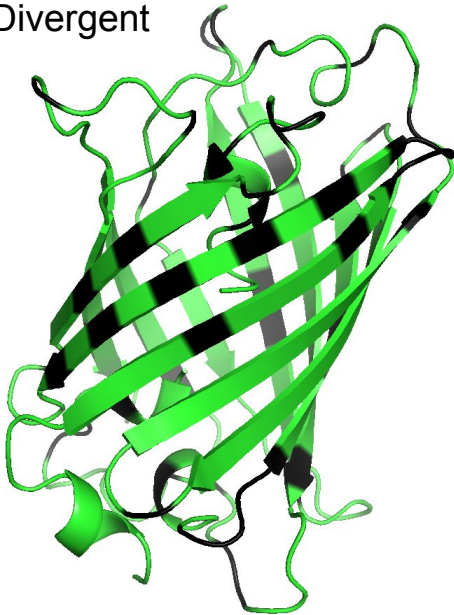


Pellet	BioBrick ¹⁶	Name	Other names	Host eukaryote	Chromophore*	Excitation max. (nm)	Emission max. (nm)	Source ref.
 S	K1033922	meffRed	meffRFP	<i>Montipora efflorescens</i>	FDYG	560	576	2
	K592012	eforRed	eforCP	<i>Echinopora forskaliana</i>	HMYG	589	609	2
	K1033927	asPink	asCP, asFP595	<i>Anemonia sulcata</i>	cMYG	568	595	19
	K1033925	spisPink	spisCP	<i>Stylophora pistillata</i>	LKYG	560	NF	2
	K1033913	scOrange	ScroogeOrange	Synthetic	cMYG			15
	K1033910	fwYellow	FezziwigYFP	Synthetic	LTYG	520	540	15
	K592010	amilGFP		<i>Acropora millepora</i>	FQYG	503	512	2
	K1033916	amajLime	amajCFP, amFP486	<i>Anemonia majano</i>	FKYG	458	486	18
 S	K592011	cjBlue		<i>Cnidopus japonicus</i>	cQYG	610	NF	20
 S	K1033902	meffBlue	Rtns5, NF podilloporin	<i>Montipora efflorescens</i>	cQYG	592	NF	21
	K864401	aeBlue	aeCP597	<i>Actinia equina</i>	cMYG	597	NF	22
	K592009	amilCP		<i>Acropora millepora</i>	cQYG	588	NF	2
	K1033906	tsPurple	TinselPurple	Synthetic	cMYG			15
	K1033927	gfasPurple	gfasCP	<i>Galaxea fascicularis</i>	sQYG	577	NF	2

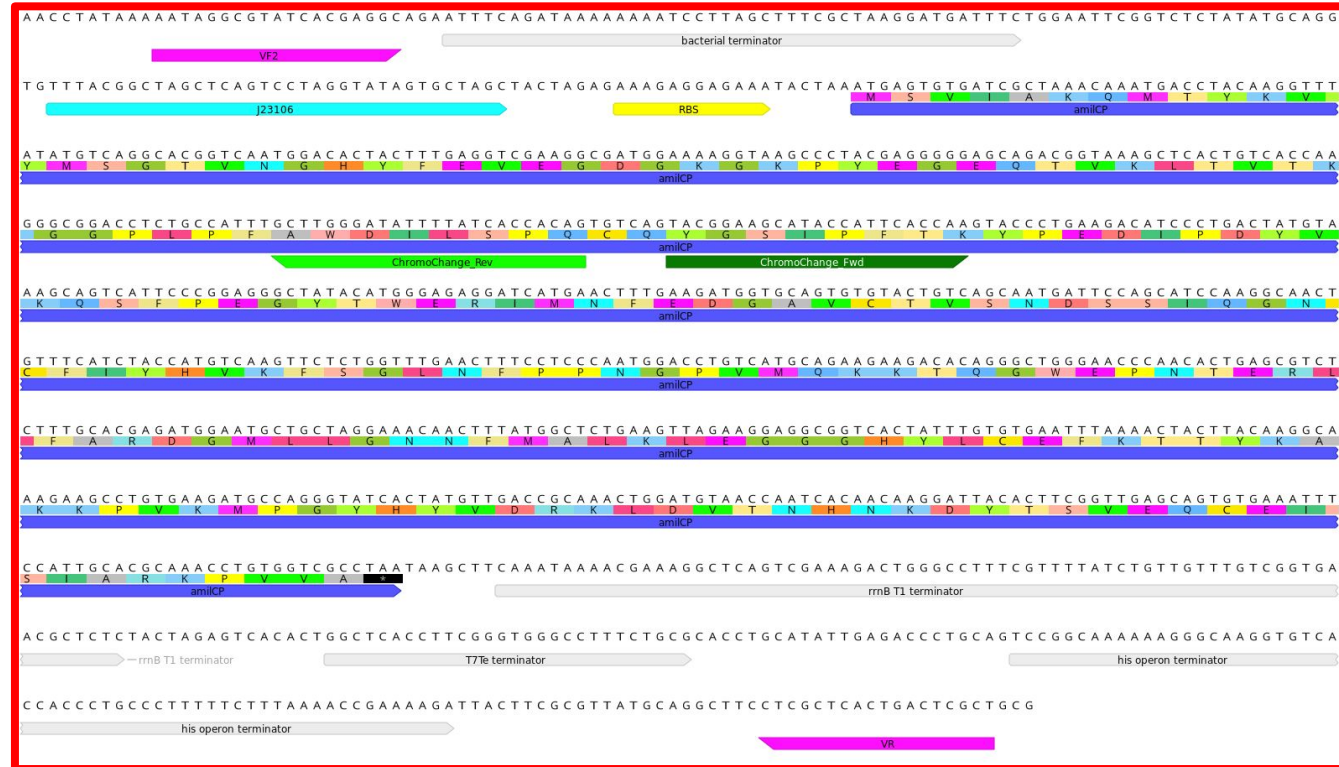
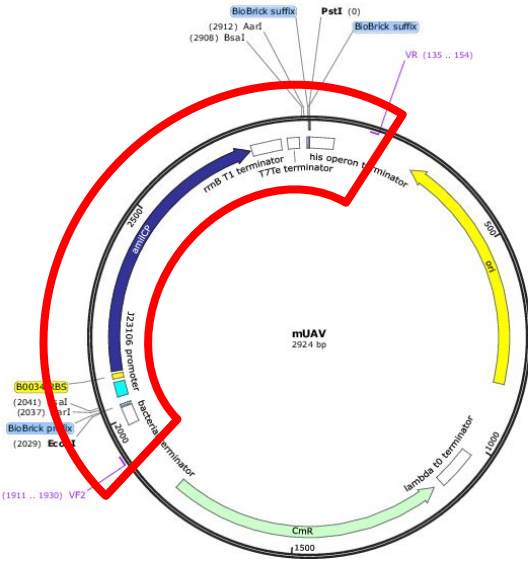
Creating the Chromophore Colorwheel



ConservedSimilarDivergent



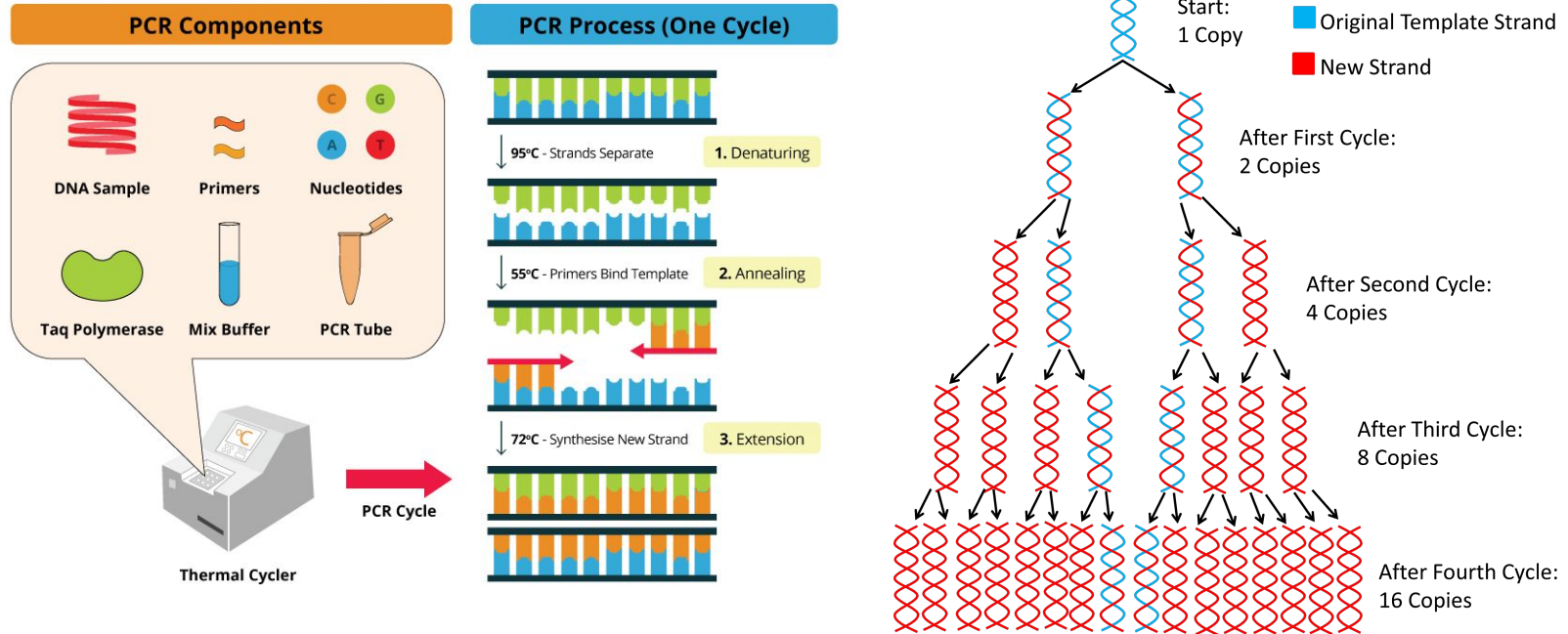
Part Map and Sequence for Assignment



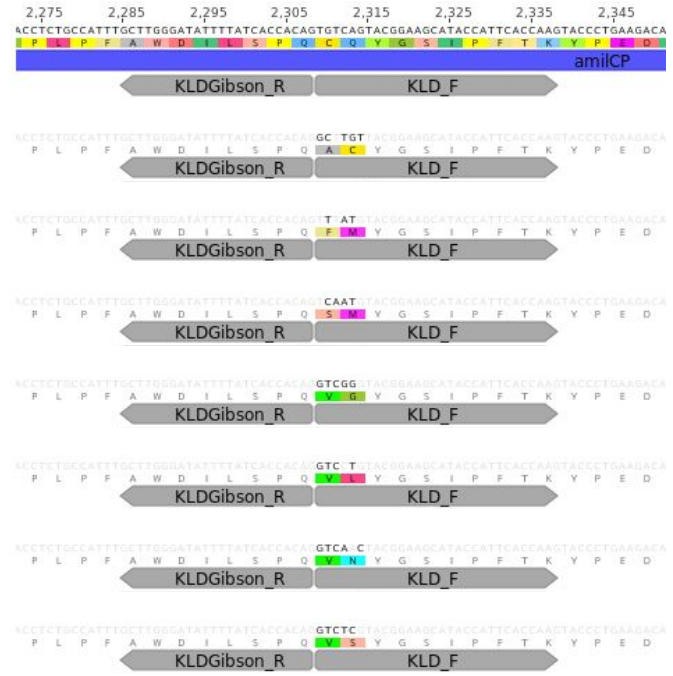
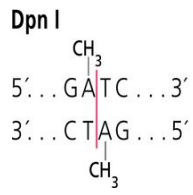
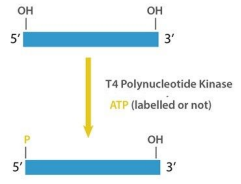
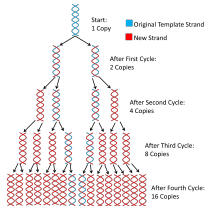
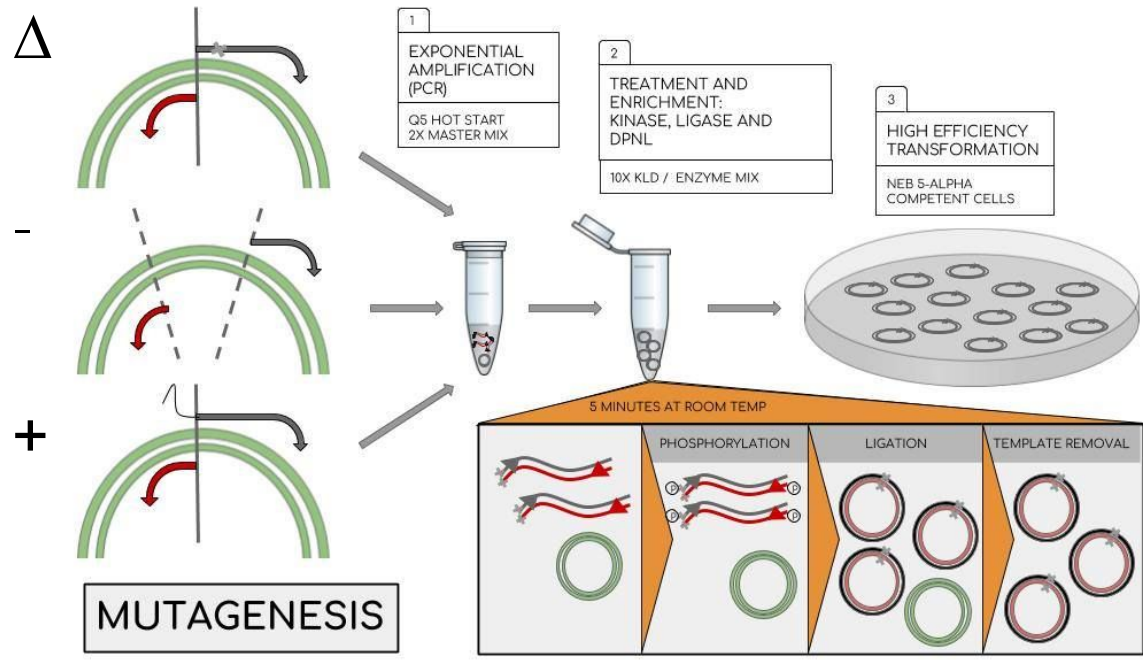
J23106 Promoter (Start DNA -> RNA)
RBS Ribosome Binding Site (RNA -> Protein)
amiCP: Chromoprotein Coding Sequence (CDS)
Terminators: Transcription Stop (Stop DNA -> RNA)

VF2: Fwd Primer Site (PCR)
VR: Rev Primer Site (PCR)

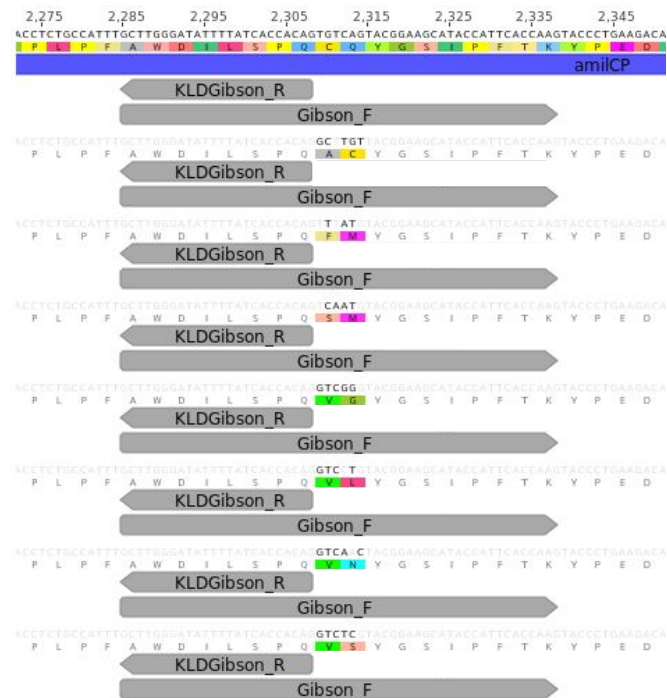
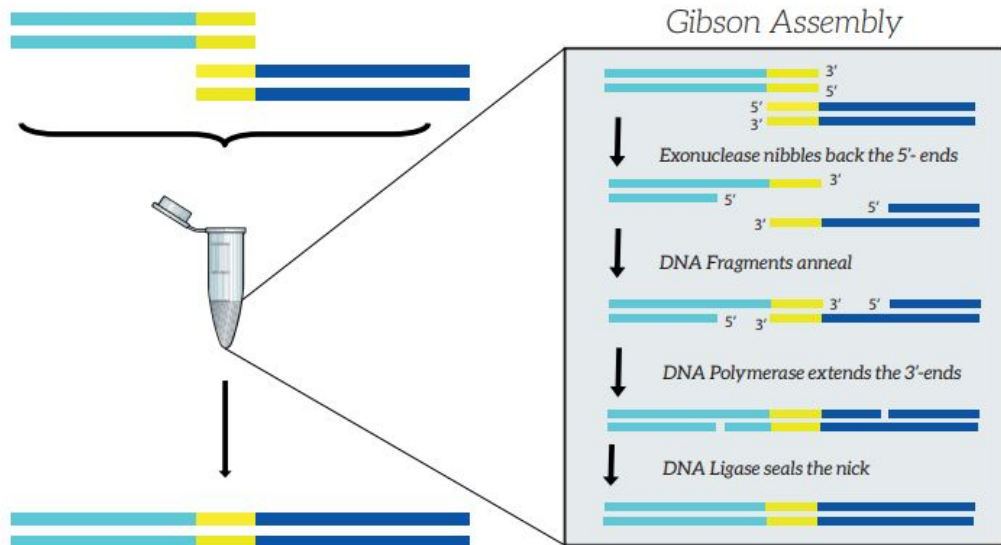
Copying and Changing DNA with Polymerase Chain Reaction



Point Mutagenesis with KLD (Kinase, Ligase,DpnI) Mix



Gibson Assembly Mechanism



NEB Gibson Assembly Cloning Kit (NEB #E5510)

- Gibson Assembly Master Mix (NEB #E2611)
- NEB 5-alpha Competent *E. coli* (NEB #C2987)

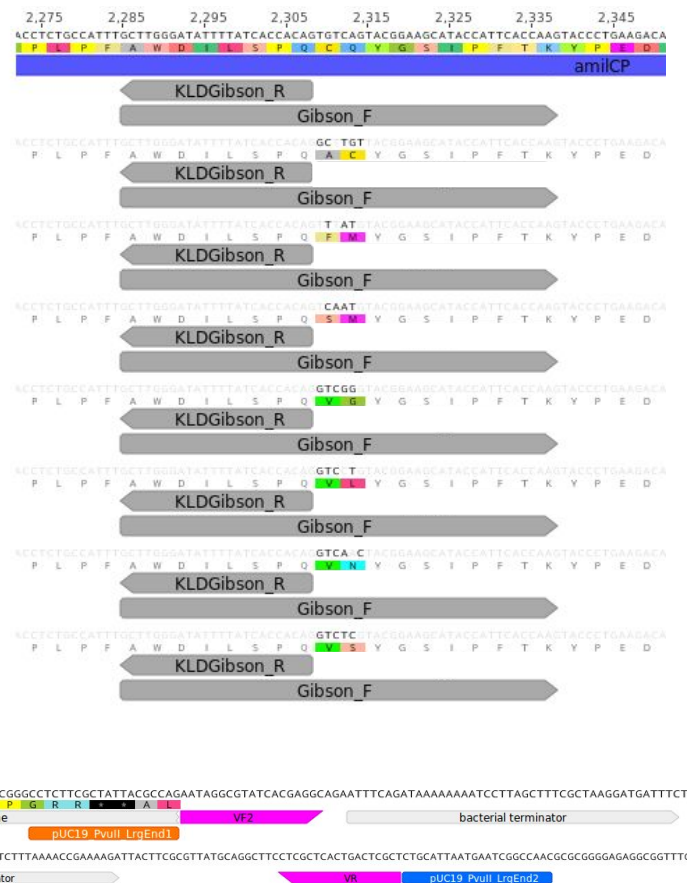
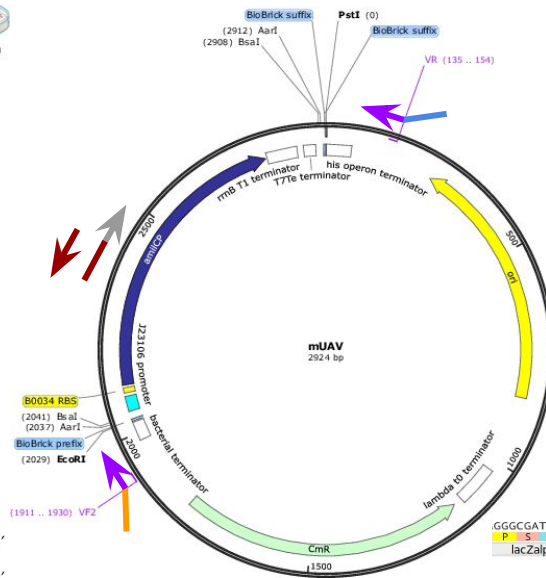
Single-tube reaction

- Gibson Assembly Master Mix
- 5' exonuclease
- DNA polymerase
- DNA ligase

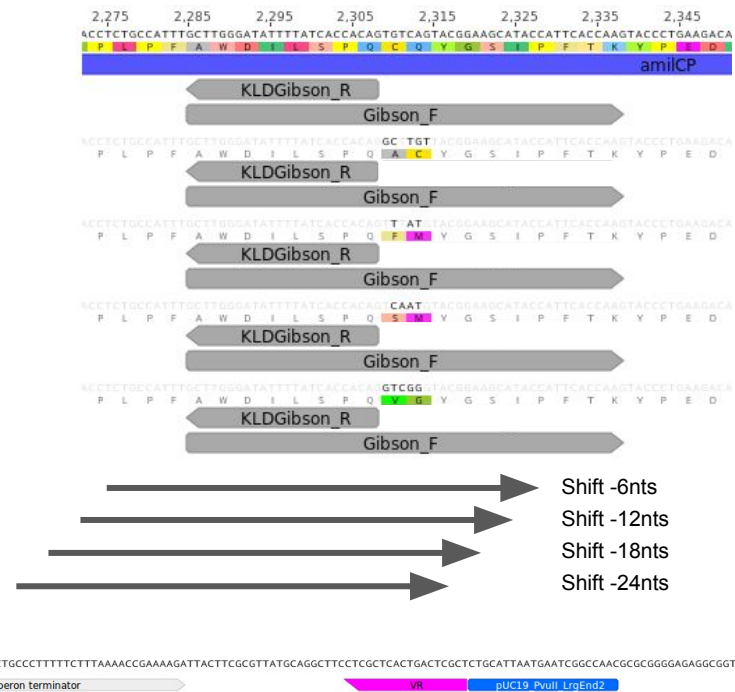
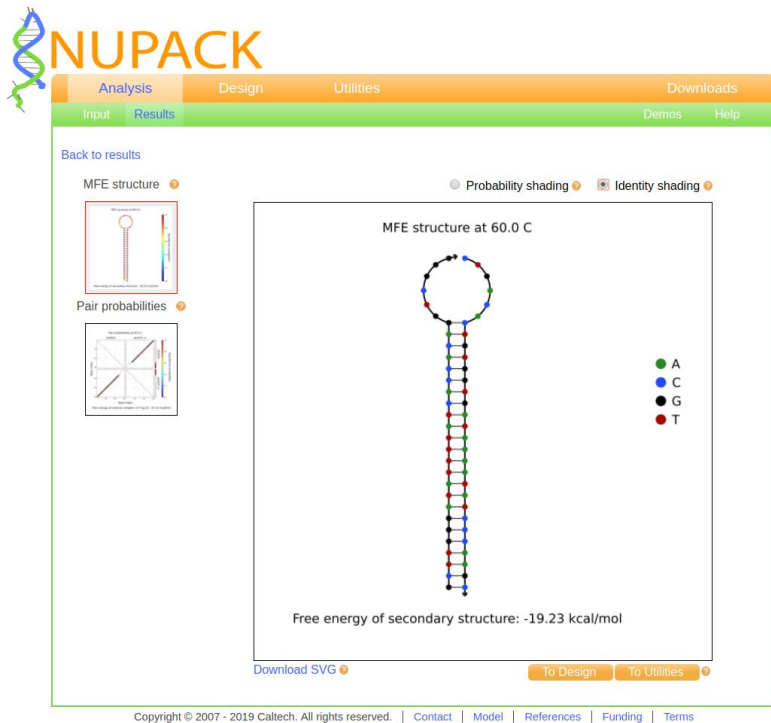
Incubate at 50°C for 15-60 minutes

Assembled DNA

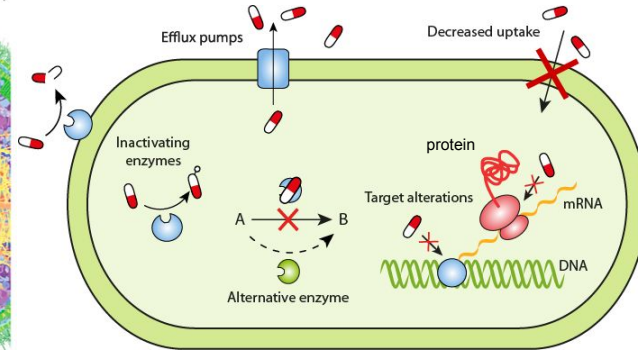
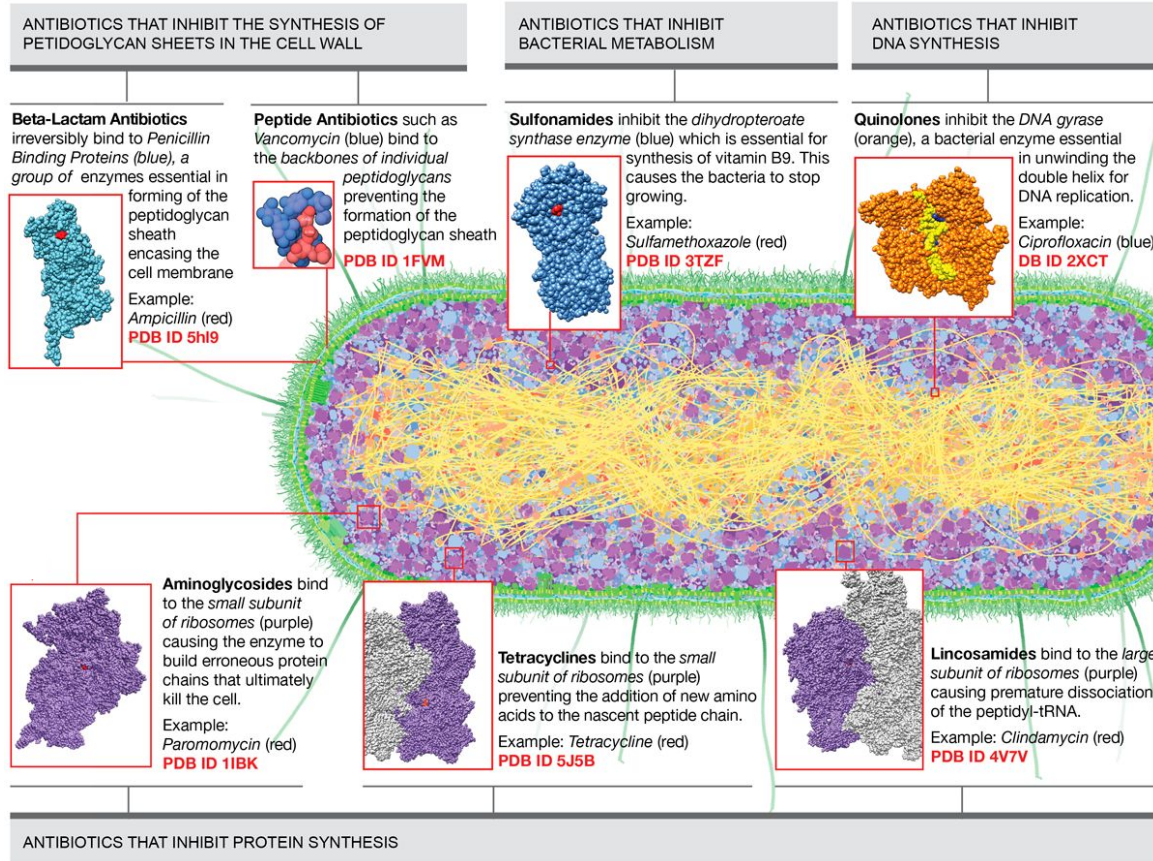
Transformation and plating



(Bonus) Primer Fitness Prediction



Alternative Antibiotic Selection



Plasmid Uptake by Calcium-Assisted Heat Shock

